

# Marine ecosystem operational services : Global Status and prospectus

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# What are « Phytoplankton Functional Types » (PFTs) ?

« Phytoplankton Functional Types » (PFTs) are conceptual groupings of several phytoplankton species, which are supposed to have in common a given ecological functionality (in terms of either the food web or the biogeochemical cycles)

For instance :

- Nitrogen fixers (e.g., *Trichodesmium*)
- Calcifiers (i.e., production of coccoliths)
- DMS producers (e.g., *Phaeocystis*)
- Silicifiers (e.g., diatoms)

The grouping is not necessarily tightly related to a physiological characteristics, but is often based on “high-level” functionalities or characteristics, such as :

- Efficient export of organic carbon to the deep ocean *versus* local recycling
- Small, medium or large sizes (pico, nano and micro- phytoplankton)

The grouping and the number of groups are related to the scientific questions that are addressed, the answer to which being supposedly better when PFTs are known



# **Why « Phytoplankton Functional Types » (PFTs) are of interest to the “biogeochemical” community ?**

- They are relevant proxies of the ecosystem functioning
- We can learn a lot about the ecosystem functioning from their time change (at all scales)
- Their respective importance will probably evolve as a function of climate change, with an impact on the efficiency of the ocean to eventually sequester carbon
- It is believed that their “incorporation” into biogeochemical models will improve the predictive capabilities of such models (a debate exists, however, as to whether or not this is really feasible and totally relevant; here we take the assertion as granted)



# Why is it suspected that PFTs are derivable from OC remote sensing ?



Both a direct and an indirect effect (not exclusive)

## Direct effect :

Changes in the phytoplankton species assemblage may lead to significant changes in the spectra of the absorption and backscattering coefficients (through changes of cell size, pigments...), which would lead to palpable changes in the reflectance spectra.

## Indirect effect :

Changes in the phytoplankton species assemblage is accompanied by a change in the ensemble of particles (detritus, viruses, bacteria...) and dissolved substances, which would lead to palpable changes in the reflectance spectra.

**If the changes of the reflectance are significant enough, they can be detected in the reflectance spectra, either *in situ* or from a remotely-sensed spectra.**

**If they are not significant enough to be directly detected, empirical relationships have to be established *in situ* between remotely-sensed quantities (e.g., Chl) and the PFTs**

# Existing techniques (1) : inversion of a reflectance model, based on a data base of forward simulations



Roesler et al., 2005.

## forward simulations

- 1 – Determine specific absorption spectra for phytoplankton species/groups
- 2 – Generate a data base of reflectance (R) spectra, for varying concentrations, compositions and size distributions
- 3 – Invert these spectra in terms of the relative contribution to absorption of each of the taxonomic groups

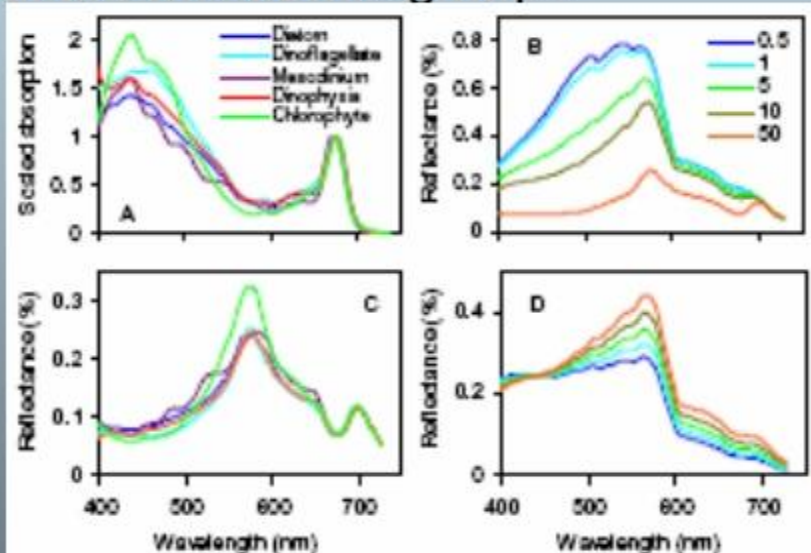


Fig. 1. (A) Scaled taxon-specific absorption spectra for 5 separable groups based upon spectrophotometric analysis of size-fractionated samples and corresponding microscopic species counts. Results of reflectance simulations as a function of (B) algal biomass ( $\mu\text{g chl l}^{-1}$ ), (C) algal composition at  $50 \mu\text{g chl l}^{-1}$  concentrations, symbols as in part A, (D) particle size distribution, where long wavelength reflectance increases as particle size increases.

The method was tested, and is quite successful, at very high biomass ( $\sim 50 \text{ mg/m}^3$ ).

Roesler, C. S., S. M. Etheridge and G. C. Pitcher. 2004. Application of an ocean color algal taxa detection model to red tides in the Southern Benguela, pp.303-305. In: Steidinger, K. A., Landsberg, J. H., Tomas, C. R., and Vargo, G. A. [eds.]. Harmful Algae 2002. Florida Fish and Wildlife Conservation Commission, Florida Institute of Oceanography, and Intergovernmental Oceanographic Commission of UNESCO.

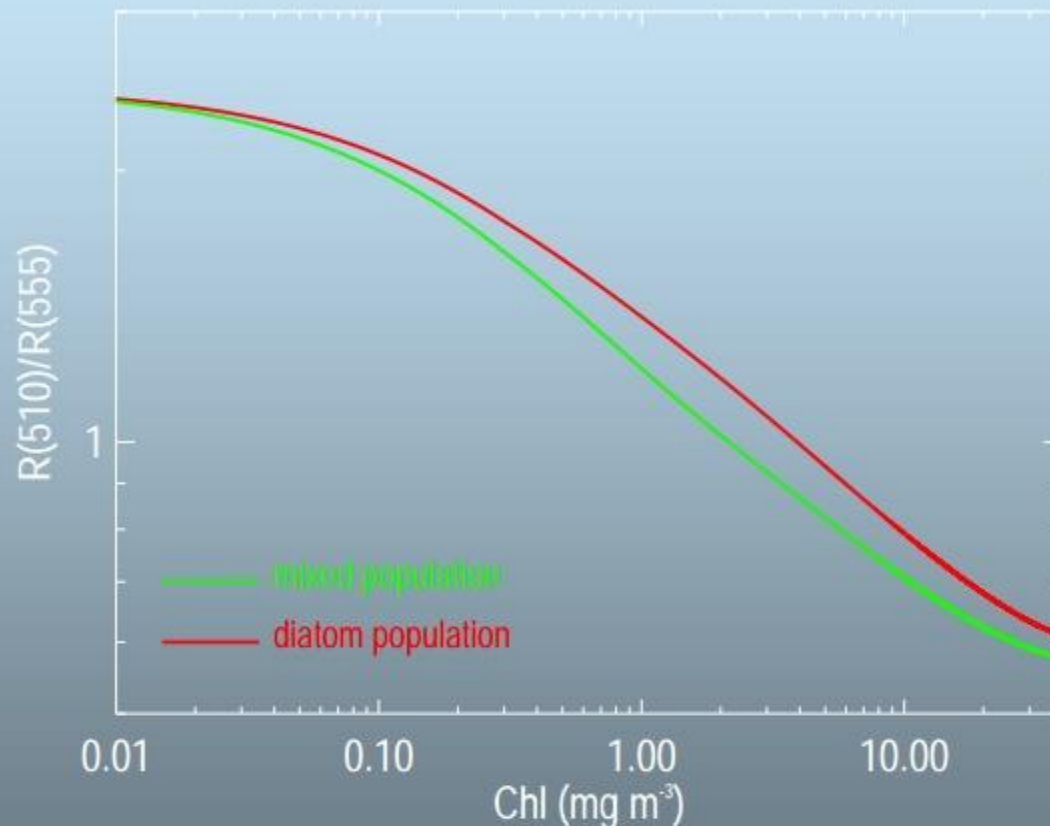


# Existing techniques (2)

E. Devred, S. Sathyendranath, C. Fuentes-Yaco, T. Platt, H. Mass  
Dalhousie Univ. & Bedford Institute of Oceanography, Canada



## Reflectance ratio as a function of chlorophyll concentration



*From S. Sathyendranath et al. 2004*

Same LUT was generated for  $R(490)/R(670)$

# Existing techniques (2, cont'd)



## Application to the Northwest Atlantic

-nLw  
:490,510,555,670 nm  
→ nRw

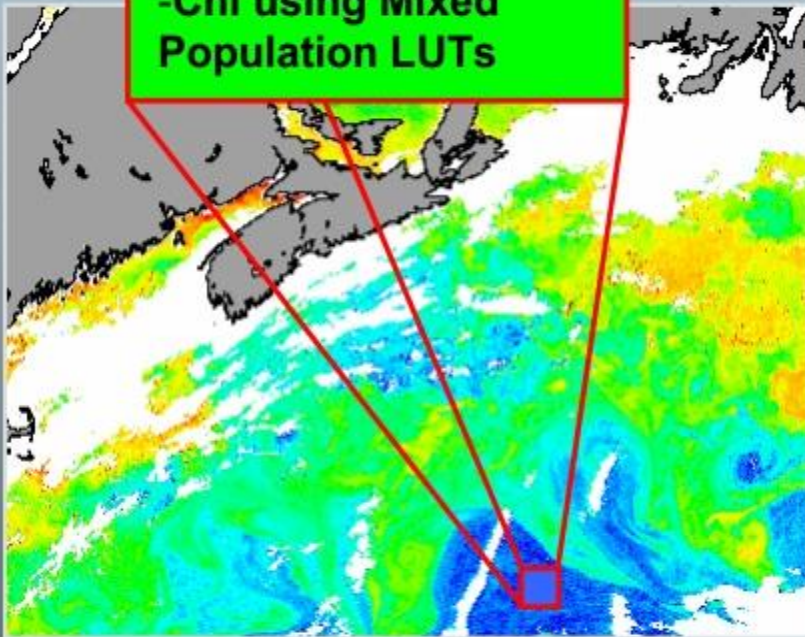
-Chl using Diatom  
LUTs

-Chl using Mixed  
Population LUTs

$$\text{Chl}_{\text{di}}(R_{510}/R_{555}) - \text{Chl}_{\text{di}}(R_{490}/R_{670}) = \varepsilon_{\text{di}}$$

$$\text{Chl}_{\text{mp}}(R_{510}/R_{555}) - \text{Chl}_{\text{mp}}(R_{490}/R_{670}) = \varepsilon_{\text{mp}}$$

$\text{Min}(\varepsilon_{\text{di}}, \varepsilon_{\text{mp}})$  yields **phytoplankton population**  
and chlorophyll concentration



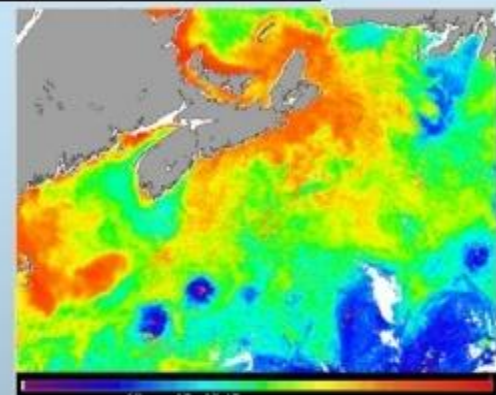
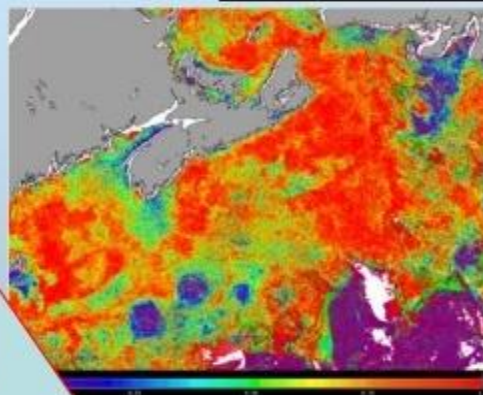


# Existing techniques (2, cont'd)

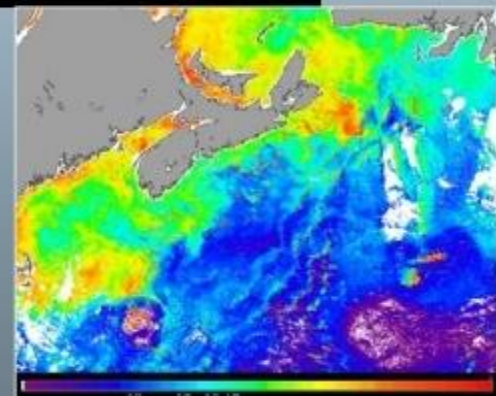
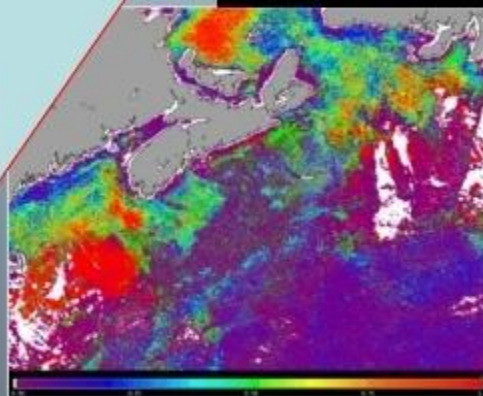


## Composite images

April 1-15 2000 (30 images)



Aug. 1-15 2000 (32 images)



Probability of occurrence  
of diatom populations

Chlorophyll concentration

$$F_{di} = \frac{\sum_n p_{di}}{\sum_n p_{di} + \sum_n p_{mp}}$$

*From S. Sathyendranath et al. 2004*

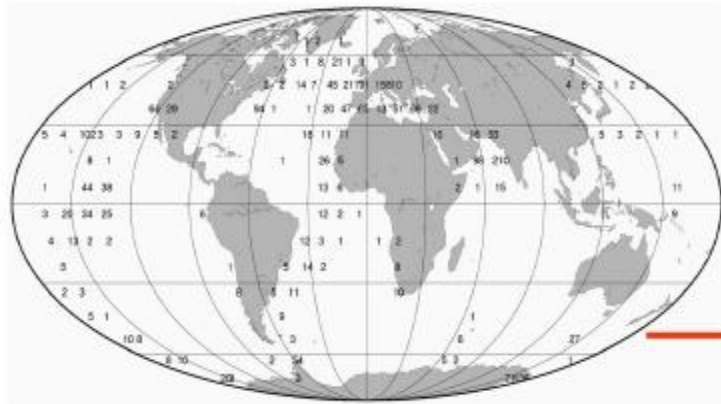


# Existing techniques (3) : relationships between surface chlorophyll, the trophic status and size classes



J. Uitz and H. Claustre, 2005

- HPLC pigment database: 2419 stations sampled between 1990 and 2002



Number of stations per square of 10°x 10°

- From diagnostic pigment (DP) to phytoplankton size classes

$$wDP = \underbrace{1.4 \text{ Fuco} + 1.4 \text{ Peri}}_{\sim \text{Micro} > 20 \mu\text{m}} + \underbrace{1.3 \text{ 19'-HF} + 0.4 \text{ 19'-BF} + 0.6 \text{ Allo}}_{\sim \text{Nano } 2-20 \mu\text{m}} + \underbrace{0.9 \text{ Zea} + 1.0 \text{ TChlb}}_{\sim \text{Pico} < 2 \mu\text{m}}$$

## Size classes proportion

$$pMicro = (1.4 \text{ Fuco} + 1.4 \text{ Peri}) / wDP$$

$$pNano = (1.3 \text{ 19'-HF} + 0.4 \text{ 19'-BF} + 0.6 \text{ Allo}) / wDP$$

$$pPico = (0.9 \text{ Zea} + 1.0 \text{ TChlb}) / wDP$$

## TChla associated to size classes: sc-TChla

$$\text{Micro-TChla} = pMicro * TChla$$

$$\text{Nano-TChla} = pNano * TChla$$

$$\text{Pico-TChla} = pPico * TChla$$

After : Claustre (L&O, 1994); Vidussi et al. (JGR, 2001)

- Standardization of the sc-TChla profiles

$$zeta = z / Z_{eu}$$

$$sc-TChla(zeta) = sc-TChla(z) / TChla_{Zeu}$$

- Interpolation of the dimensionless sc-TChla profiles

$$zeta = 0 \text{ (surface)} \text{ to } zeta = 2 \text{ (} 2 * Z_{eu} \text{)} \Rightarrow 20 \text{ points / profile}$$

- Sorting of the interpolated sc-TChla profiles

$\Rightarrow$  according to the hydrological regime: stratified / mixed

$\Rightarrow$  according to  $[TChla]_{surf}$

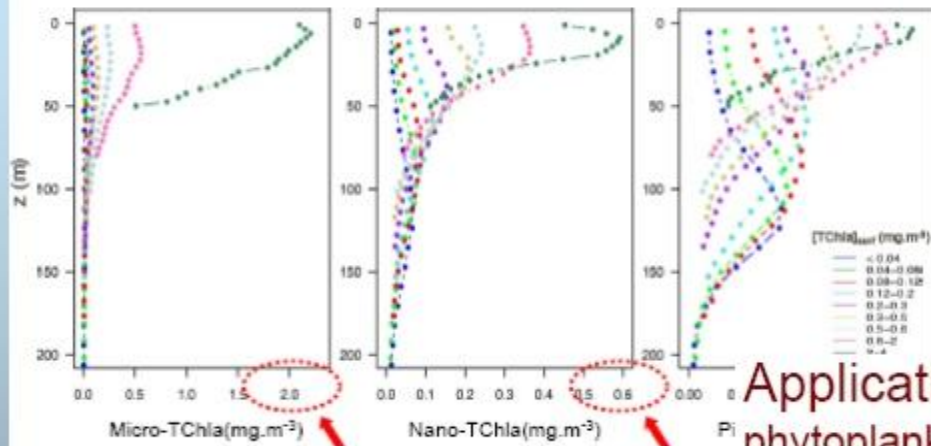
- Computation of average sc-TChla profiles / trophic class

After : Morel & Bérthon (L&O, 1989)

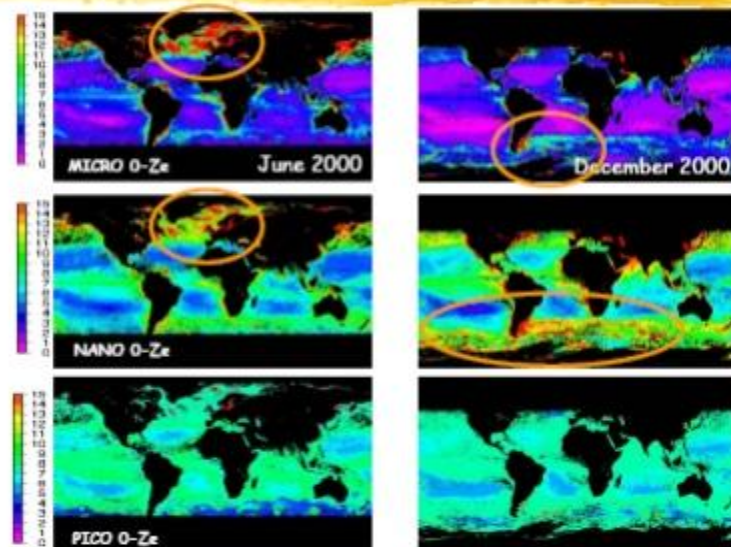
# Existing techniques (3, cont'd)



## Geometrically rescaled profiles



Application:  
phytoplankton functional groups climatology



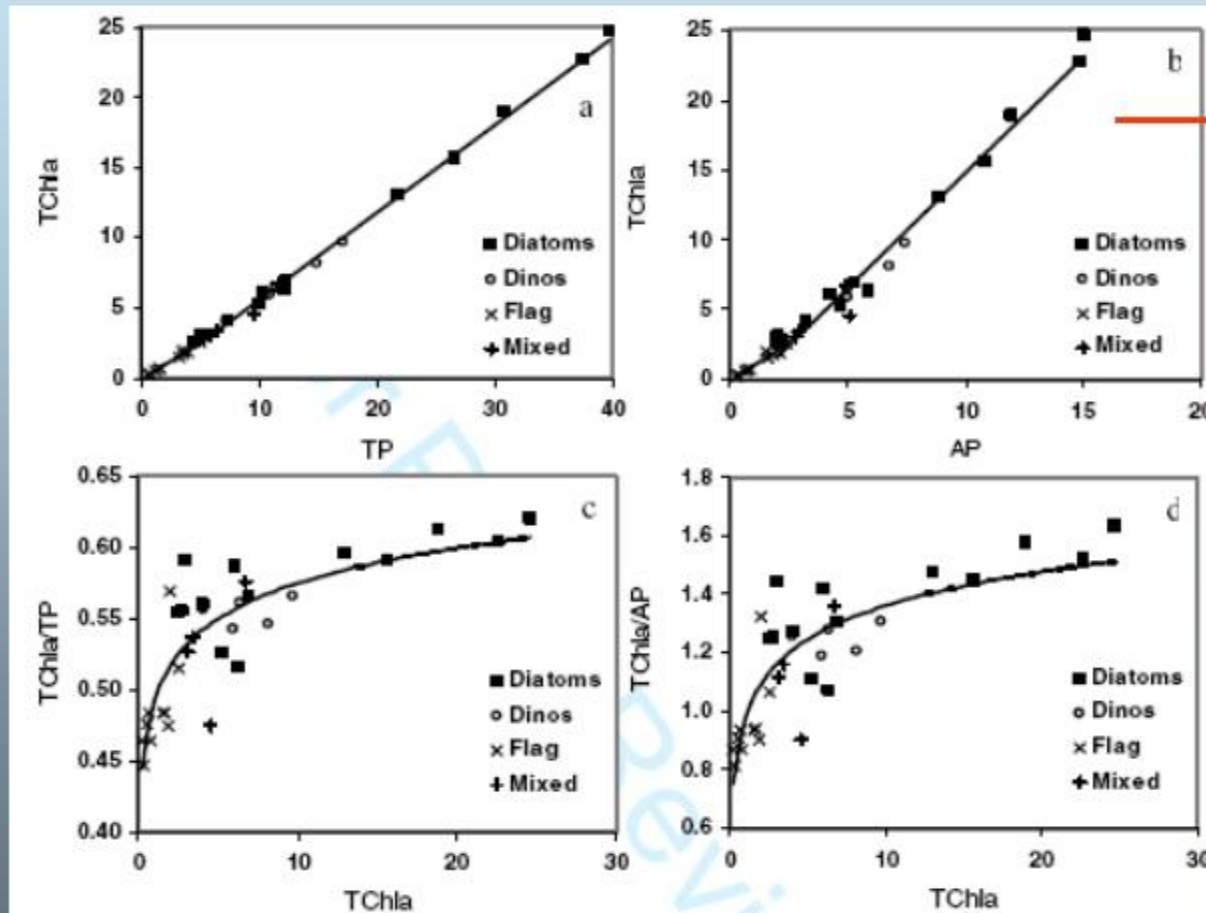


# Existing techniques (4) :



Aiken *et al.*, 2005

HPLC-determined pigments are used to distinguish between groups



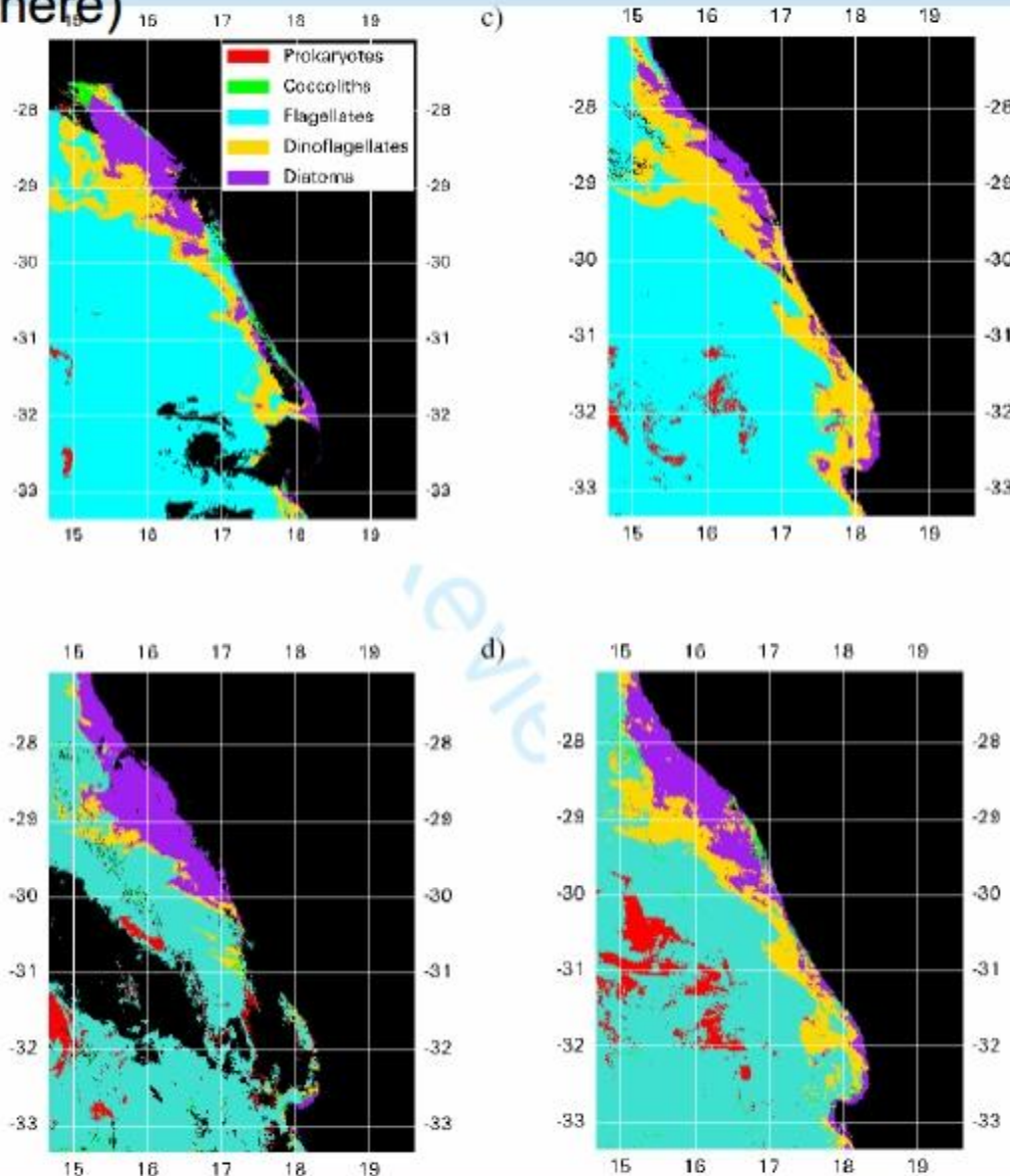
Similar relationships and associated statistics are derived for several phytoplankton groups, based on a data set of inherent optical properties + HPLC pigments. Therefore, groups can be determined from Chl-*a*

Courtesy : J. Aiken

# Existing techniques (4, cont'd)



The method is then applied to satellite images (MERIS images here)



Example here : the BENCAL cruise (Sept 2002) in the Benguela upwelling

Courtesy : J. Aiken



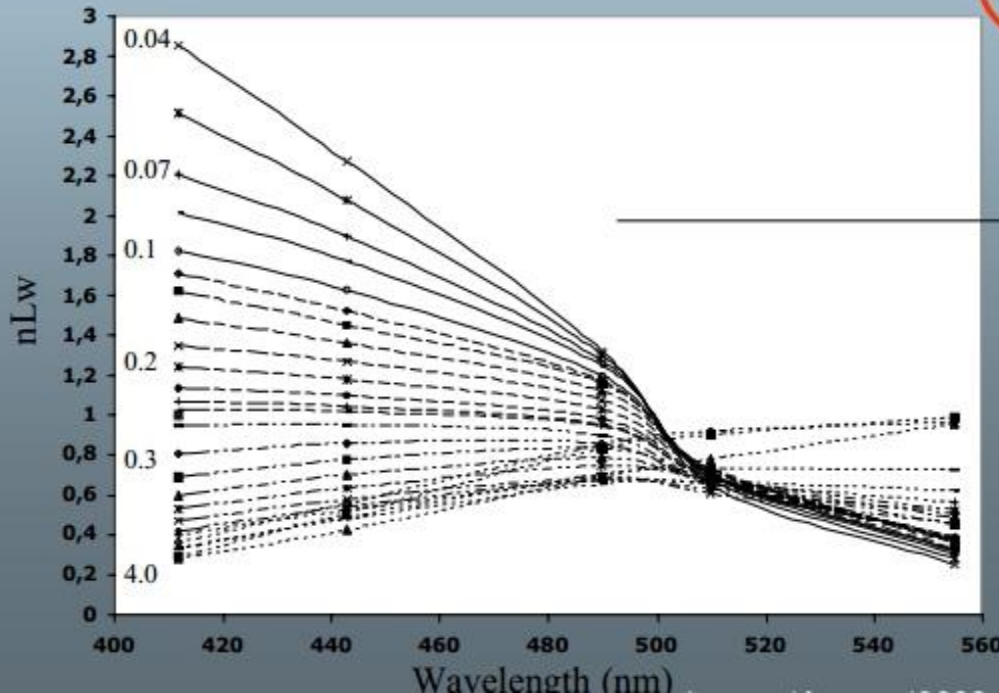
# Existing techniques (5) : empirical relationships between groups (HPLC-determined) and ocean colour



Alvain and Moulin (Deep-Sea Research I, 2005, vol , xx-xx).

- 1 – Determine the dominant group from HPLC pigment inventories (Gep&CO program)
- 2 – Put together these groups and the SeaWiFS-derived nLw's, after the 1<sup>st</sup> order effect of Chl has been removed
- 3 – Determine specific nLw\* spectra for each group
- 4 – Extend to global SeaWiFS imagery

$$nLw^*(\lambda) = nLw_{obs}(\lambda) / nLw_{mod}(Chl-a_{swfs}, \lambda)$$

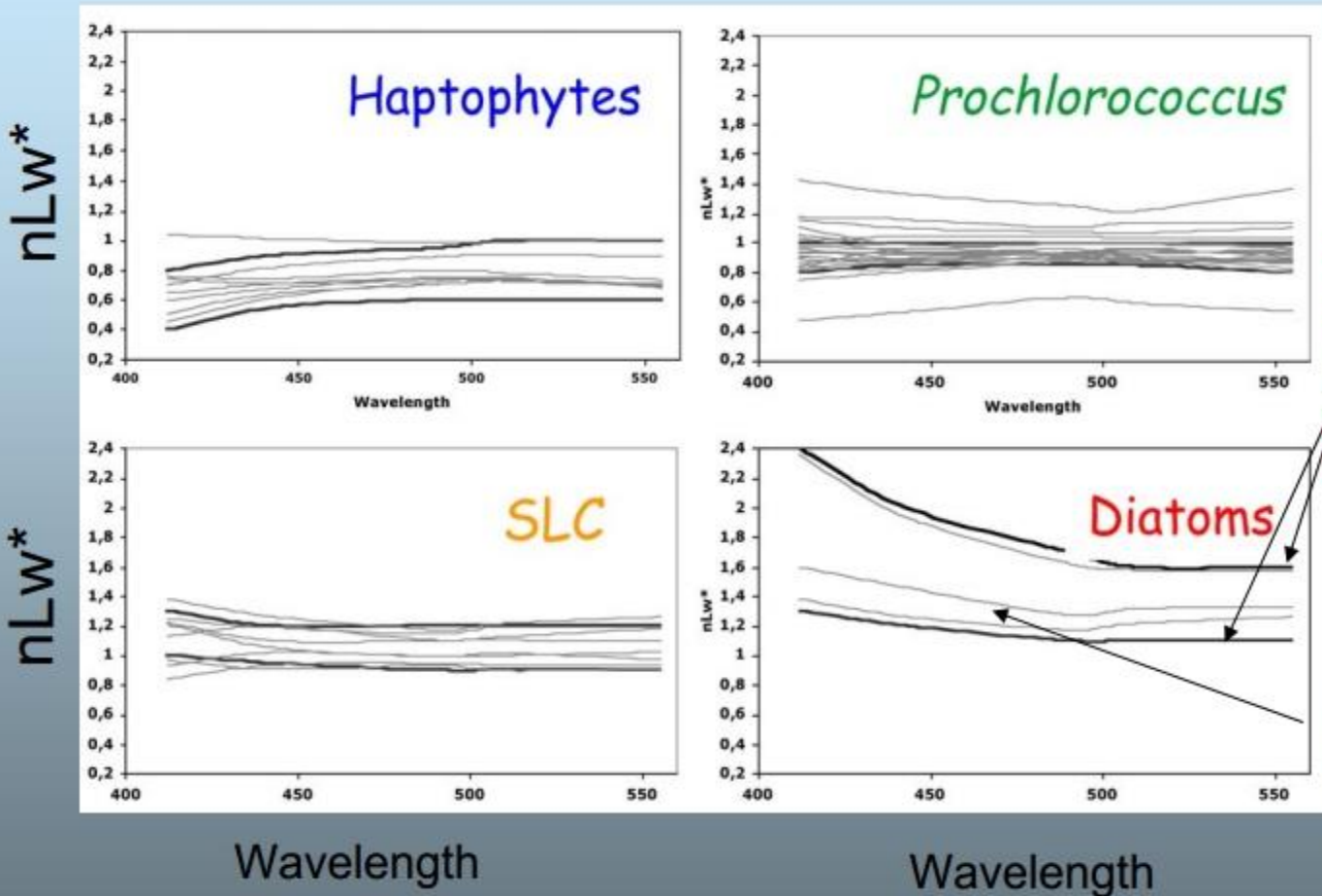


Courtesy: S. Alvain



## Existing techniques (5, cont'd)

A specific  $nLw^*$  for each dominant group !



Envelope of  $nLw^*$  spectrum  
used to characterize the phytoplankton group

Courtesy:  
S. Alvain

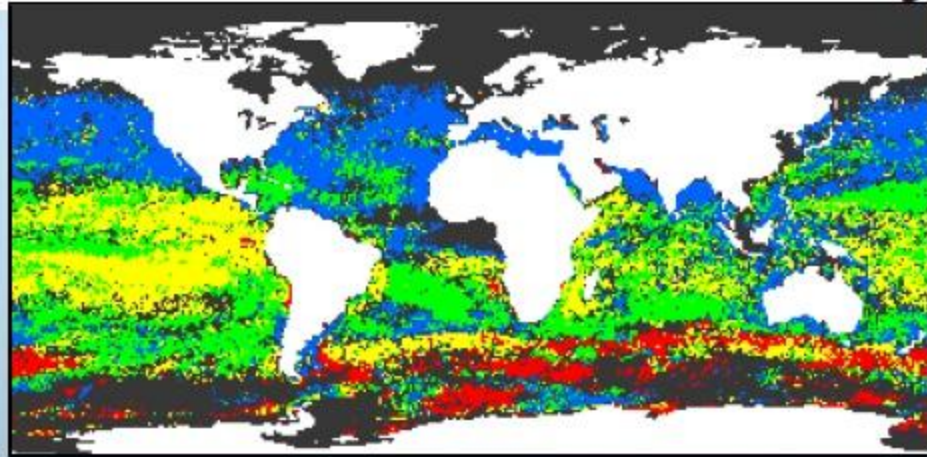
*In light grey :  
Individual spectral  
signature  
for the 41  $nLw^*$  associated  
with the 41 Gep&Co  
pigments inventories*

Is the  $nLw^*$  spectrum usable to detect dominant phytoplankton groups at the global scale ?

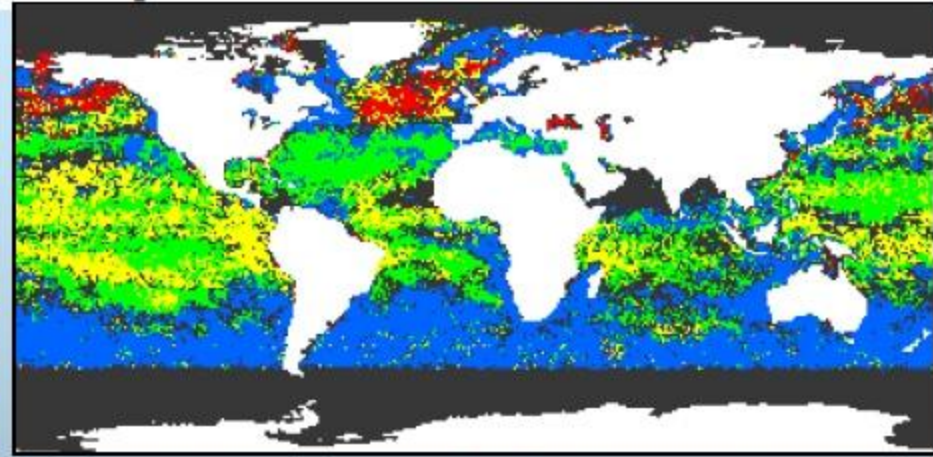


# Existing techniques (5, cont'd)

## Global monthly maps for 2001



January



April

→ Dominant groups

From Alvain *et al.*, 2005

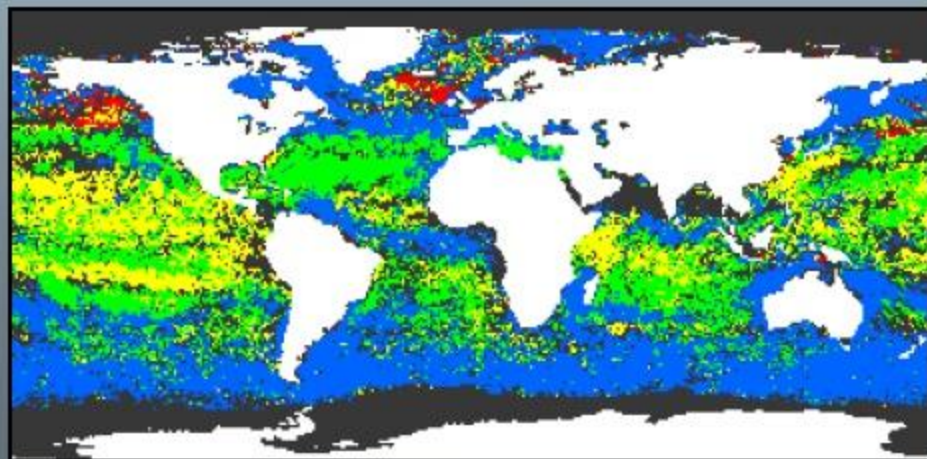
Haptophytes

*Prochlorococcus*

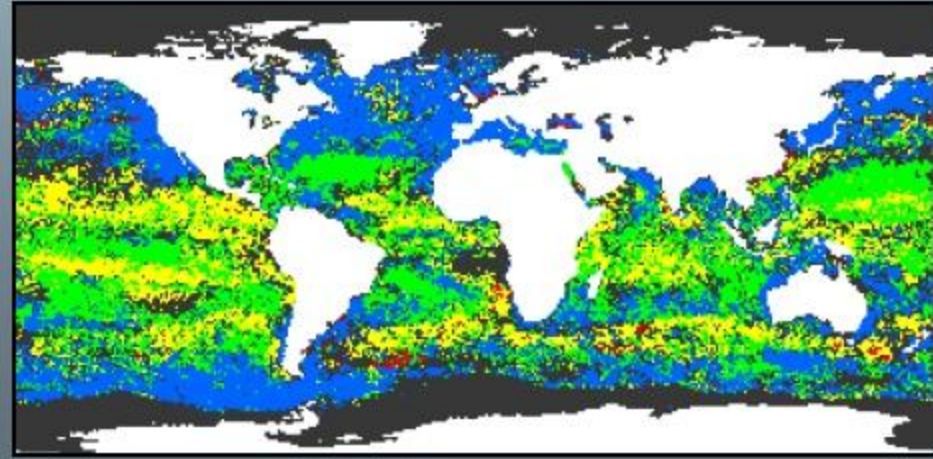
SLC

Diatoms

« PHYSAT »



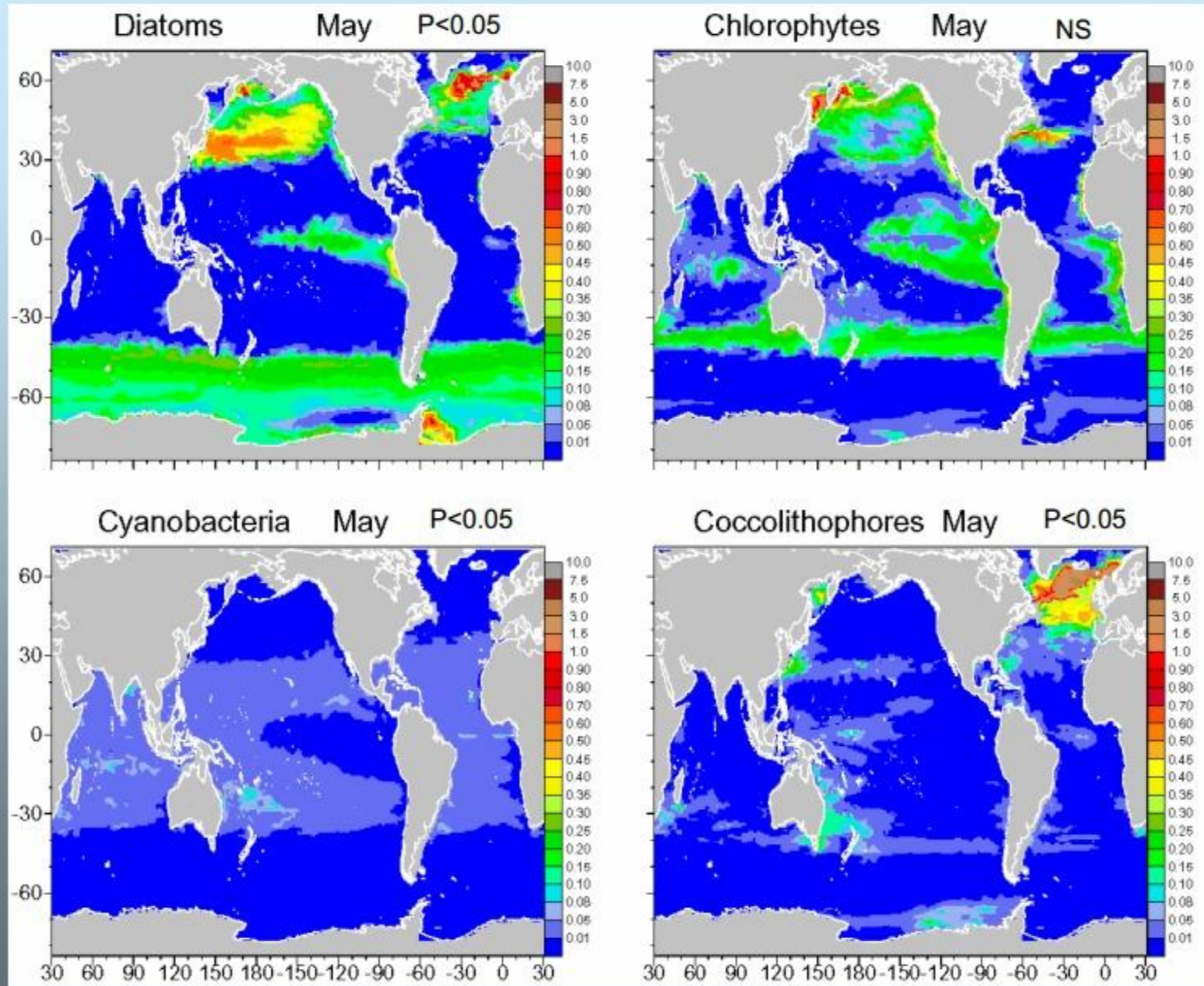
June



October



# Examples of PFTs from global models (1)



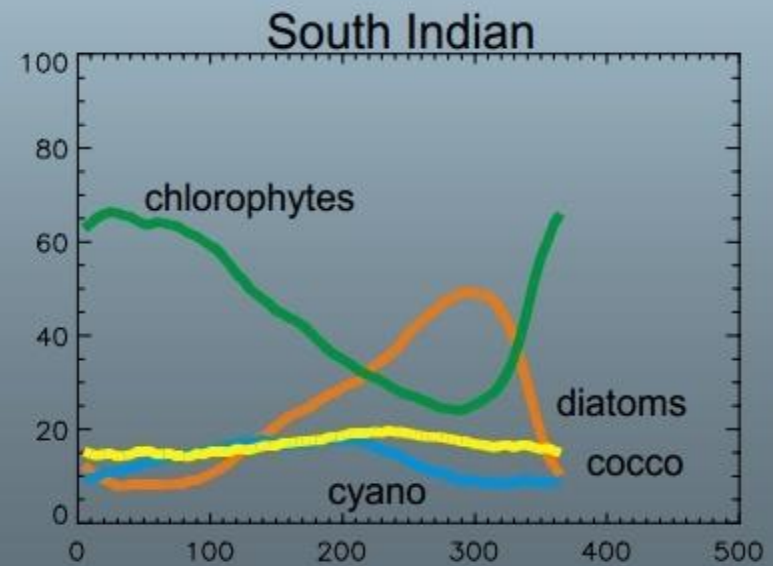
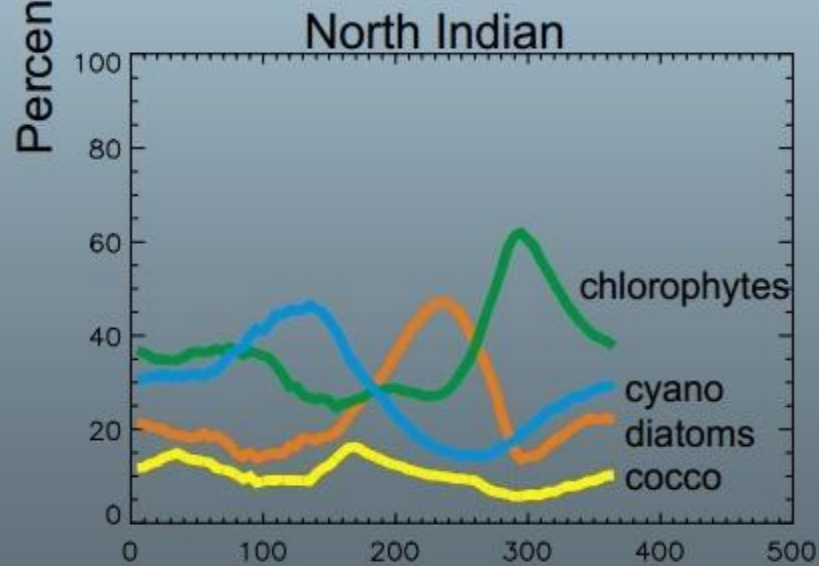
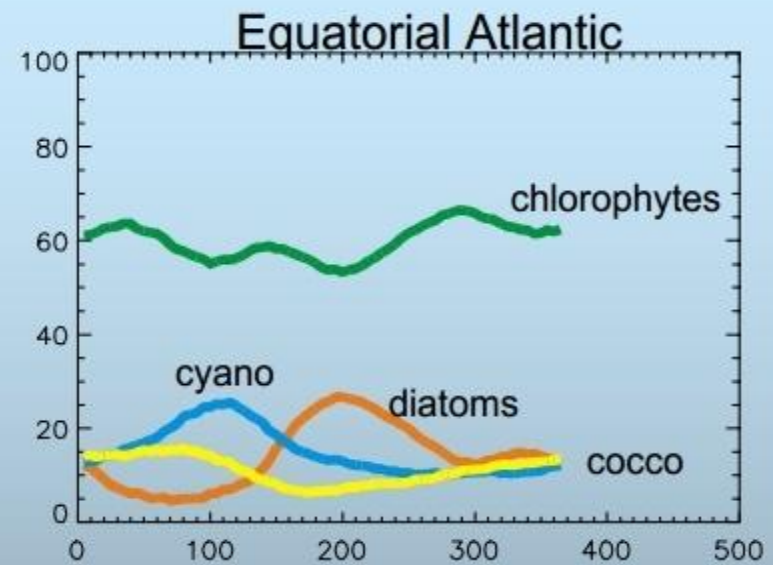
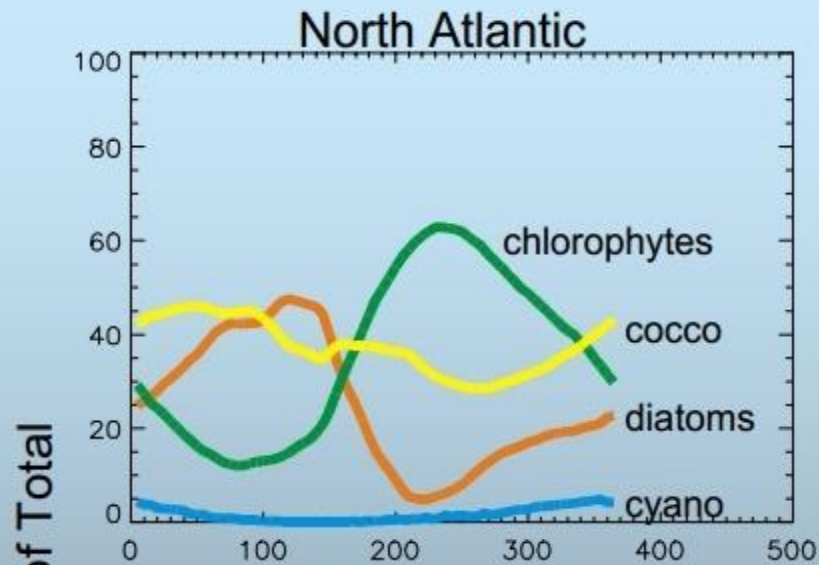
Courtesy: W. Gregg



# Examples of PFTs from global models (1, cont'd)



Courtesy: W. Gregg



Day of Year

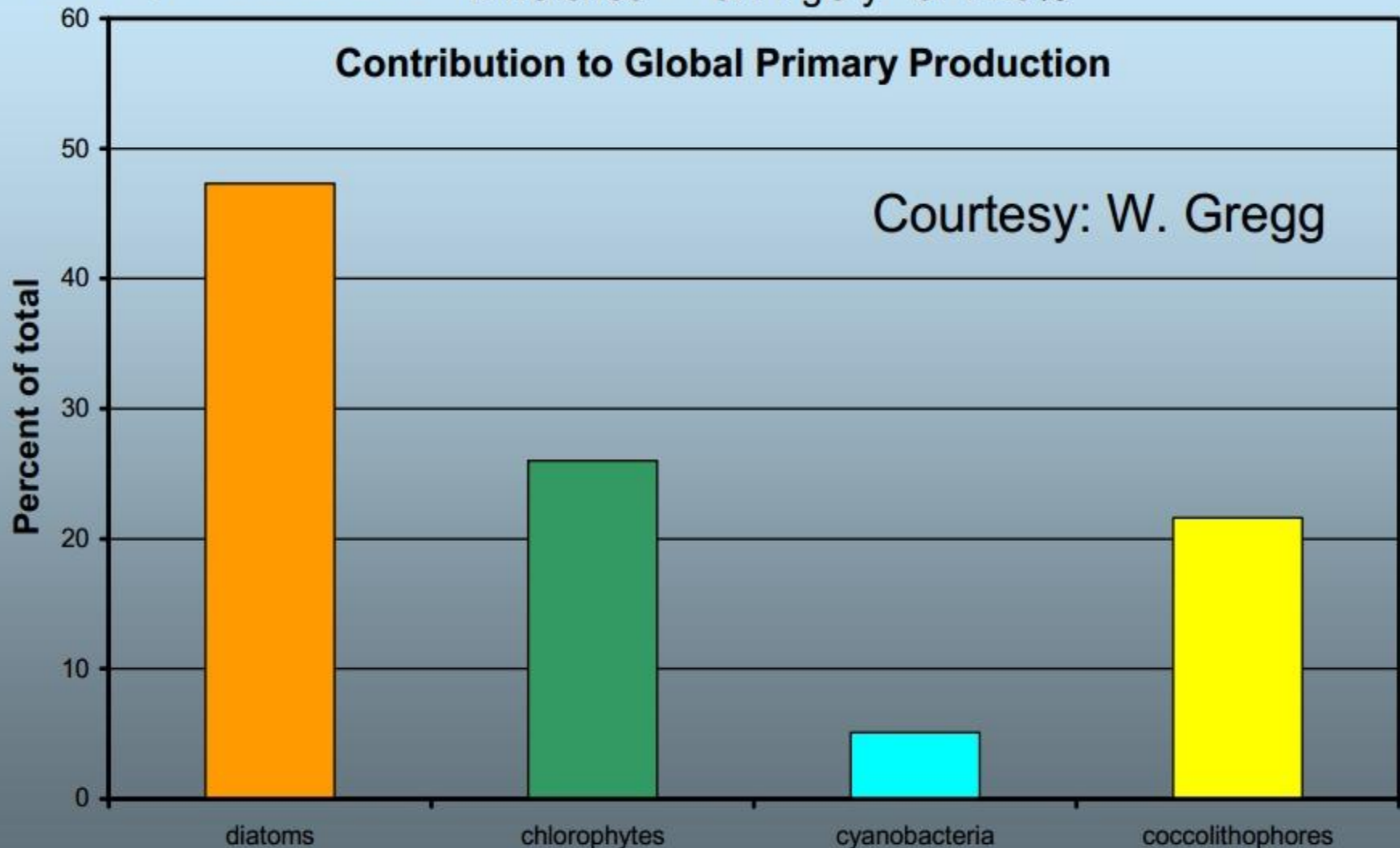
Model-estimated seasonal variability (Gregg et al., 2003)

# Examples of PFTs from global models (1, cont'd)



Model = 48.9 PgC y<sup>-1</sup>  
SeaWiFS = 42.7 PgC y<sup>-1</sup>  
Difference = 6.1 PgC y<sup>-1</sup> or 14.3%

## Primary Production



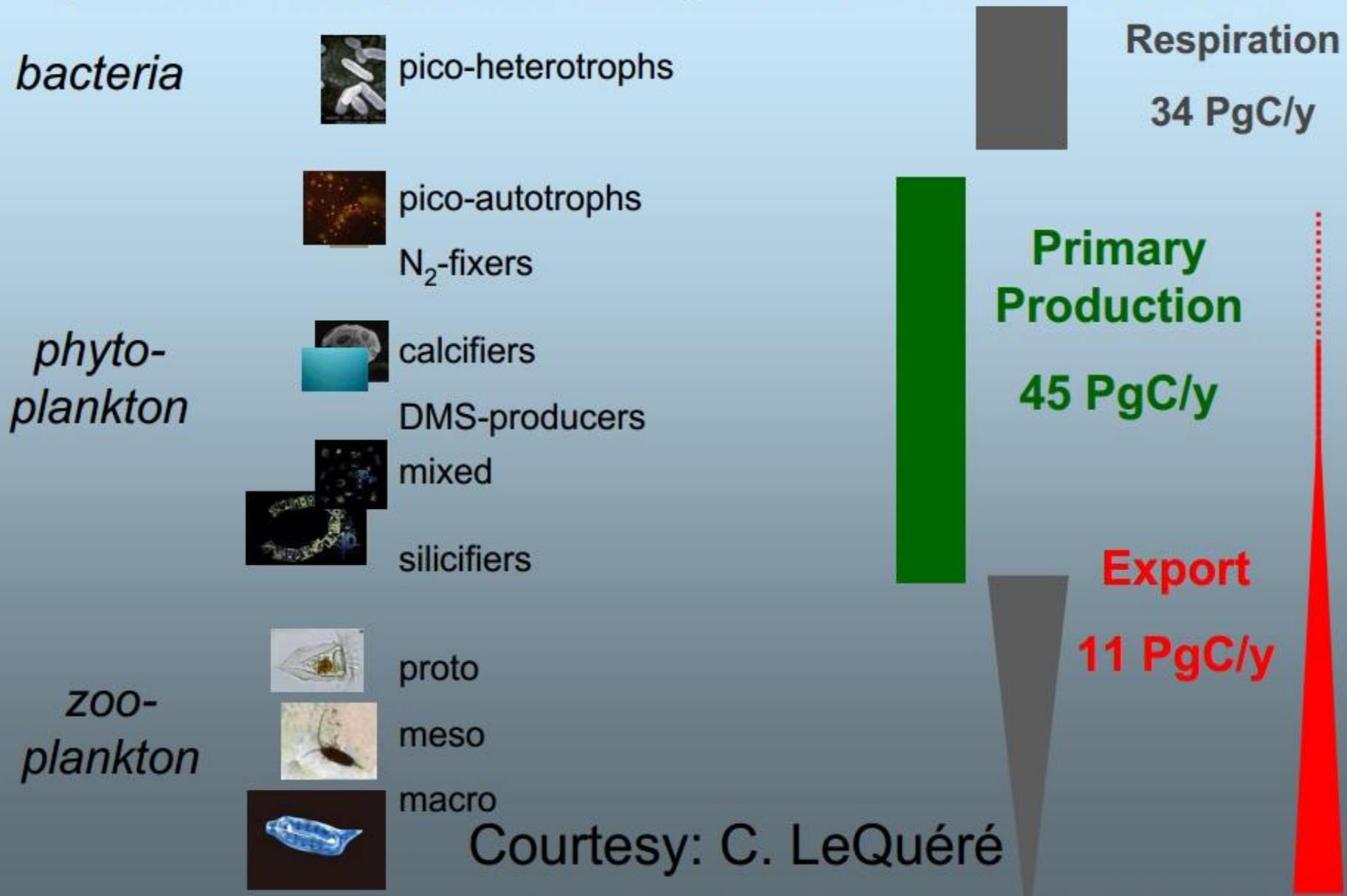
Model estimates of primary production (Gregg, 2005, unpublished)



# Examples of PFTs from global models (2)



planned developments of a Dynamic Green Ocean Model

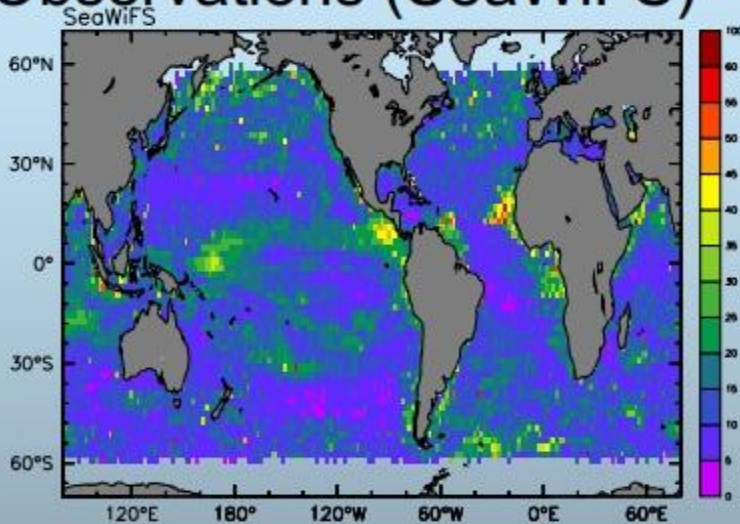


# Examples of PFTs from global models (2, cont'd)

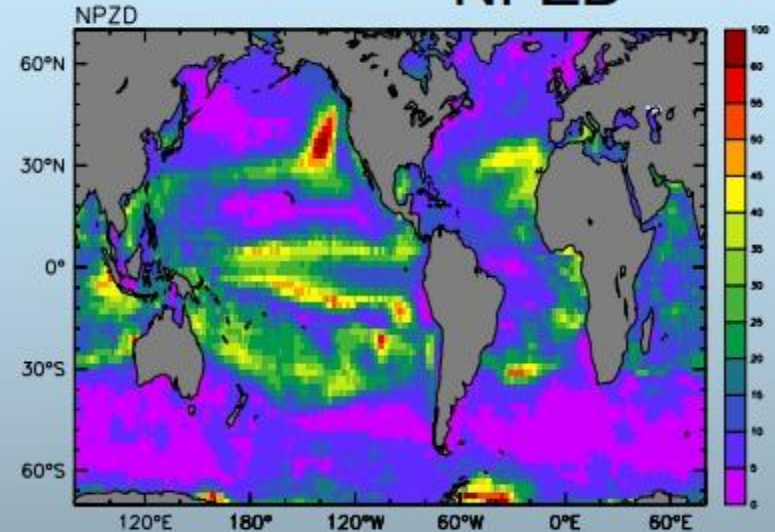


Interannual chla variability (percent)

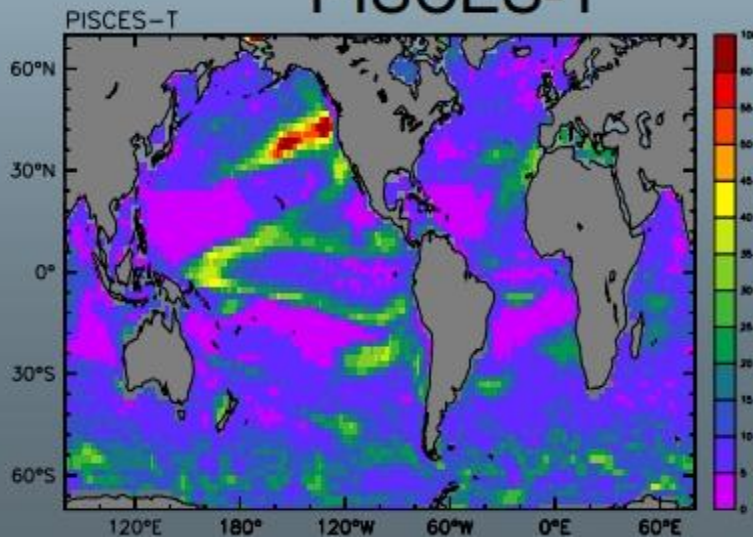
Observations (SeaWiFS)



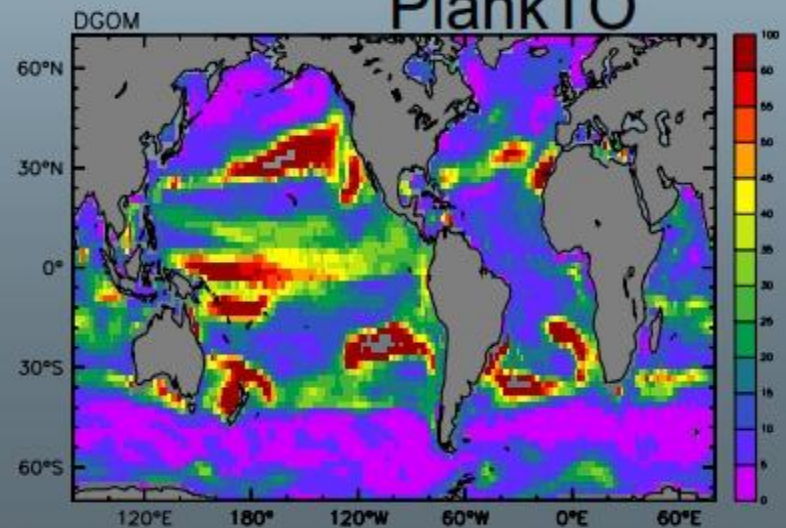
NPZD



PISCES-T



PlankTO



Courtesy: C. LeQuéré

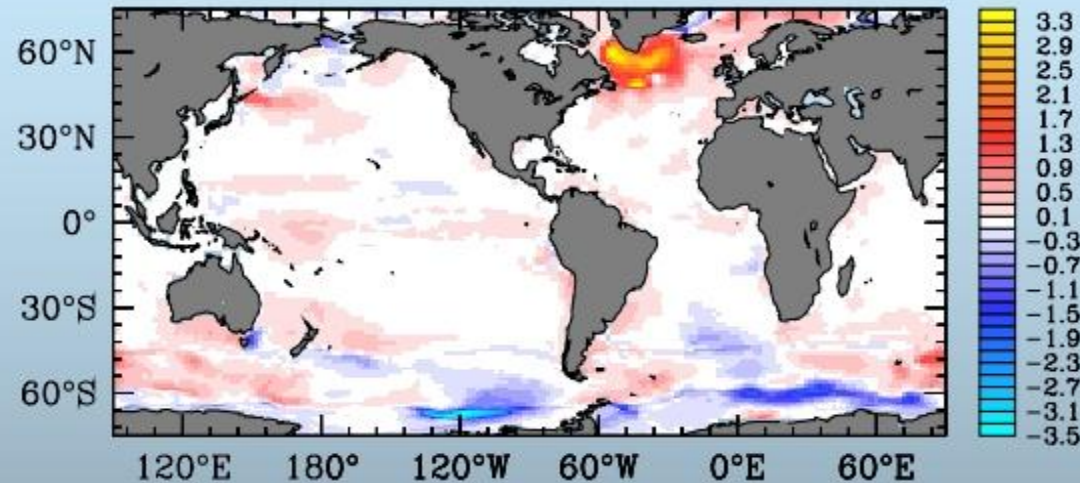
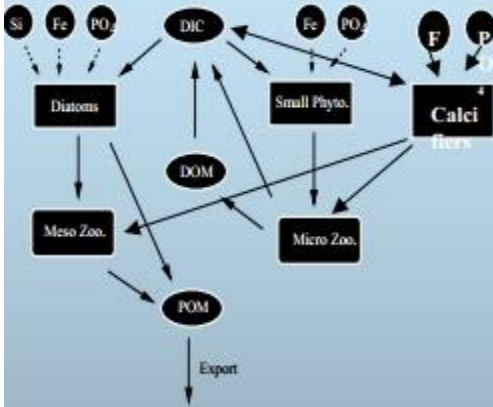


# Examples of PFTs from global models (2, cont'd)

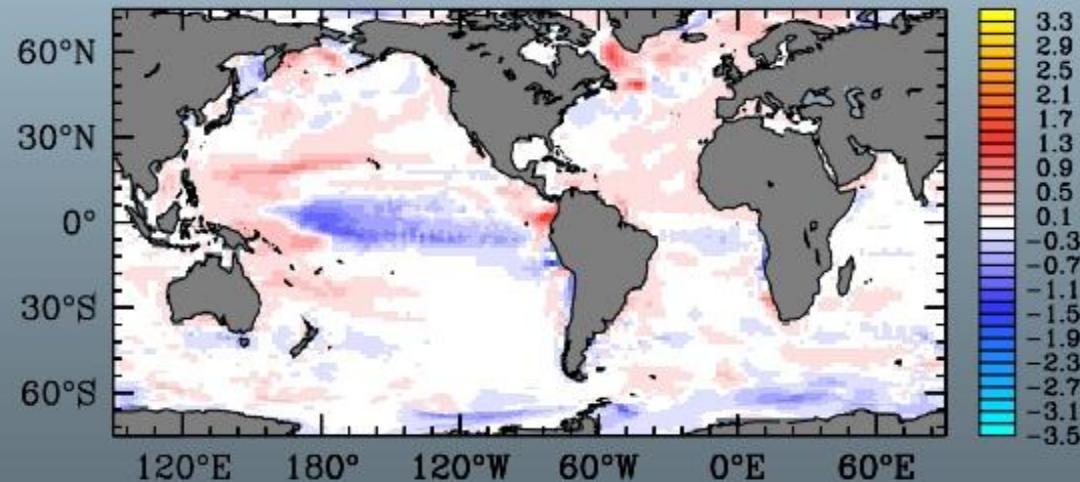
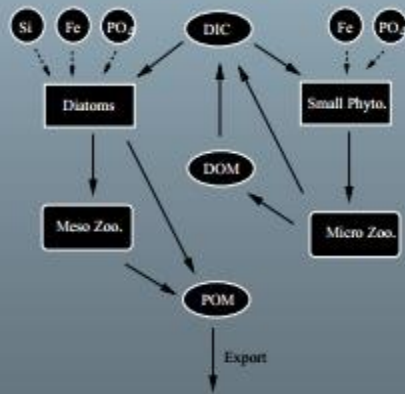


Impact of climate change on the CO<sub>2</sub> sink in 2060 using identical physics but two different ecosystem models (mol/m<sup>2</sup>/y)

## PlankTOM



## PISCES-T



Courtesy: C. LeQuéré



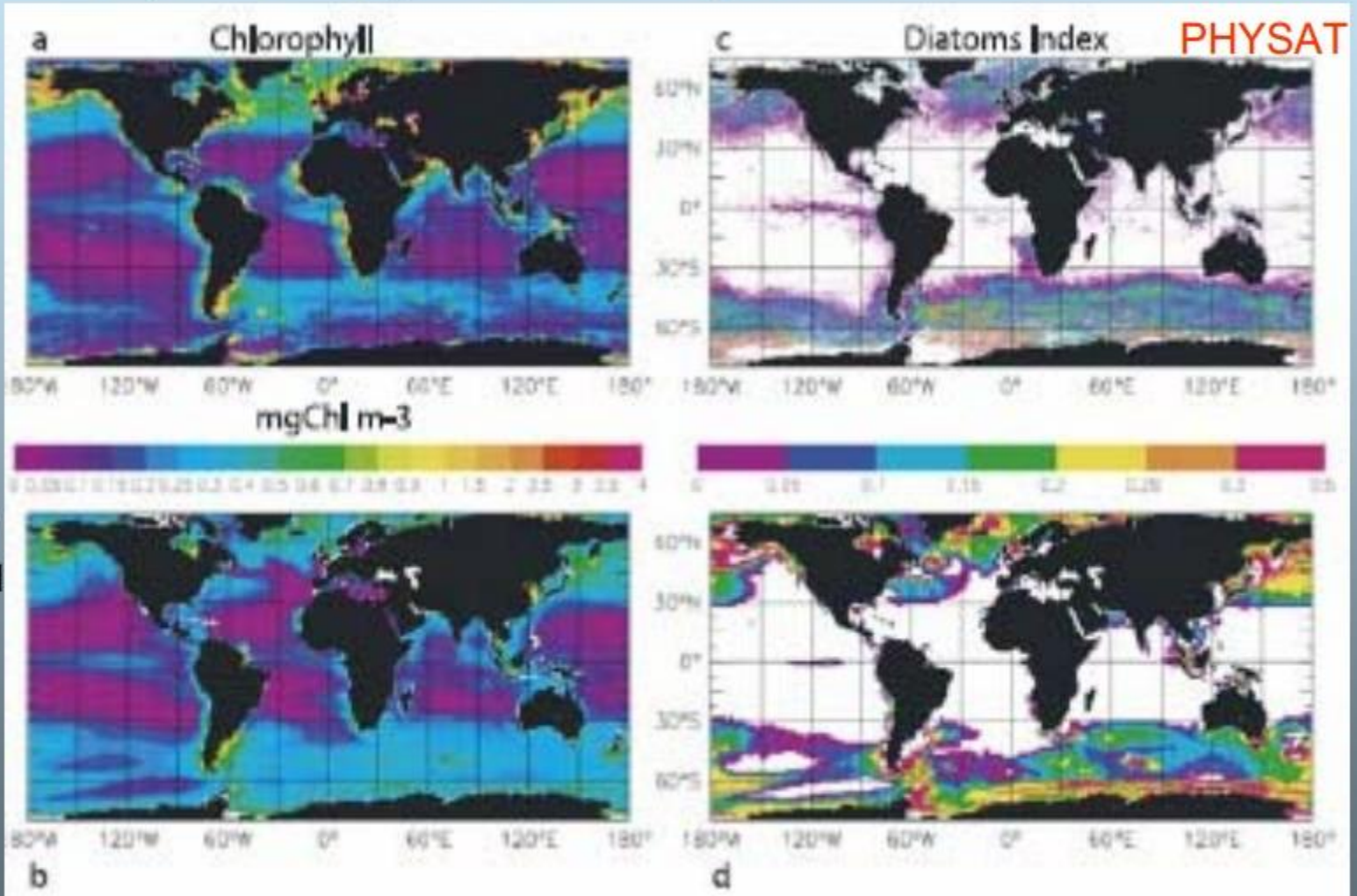
# Examples of PFTs from global models (3)



From Bopp *et al.*, 2005

“Response of diatoms distribution to global warming and potential implications: A global model study”, GRL, vol 32, 2005

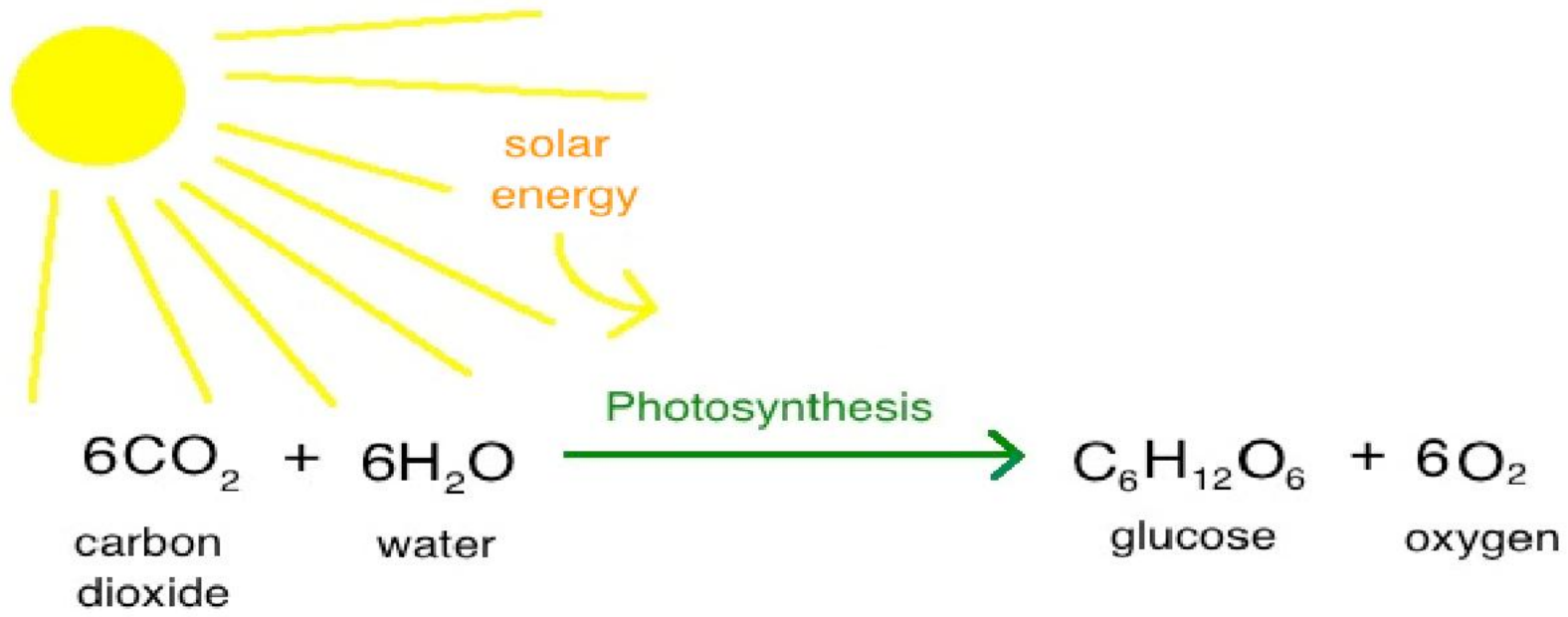
SeaWiFS



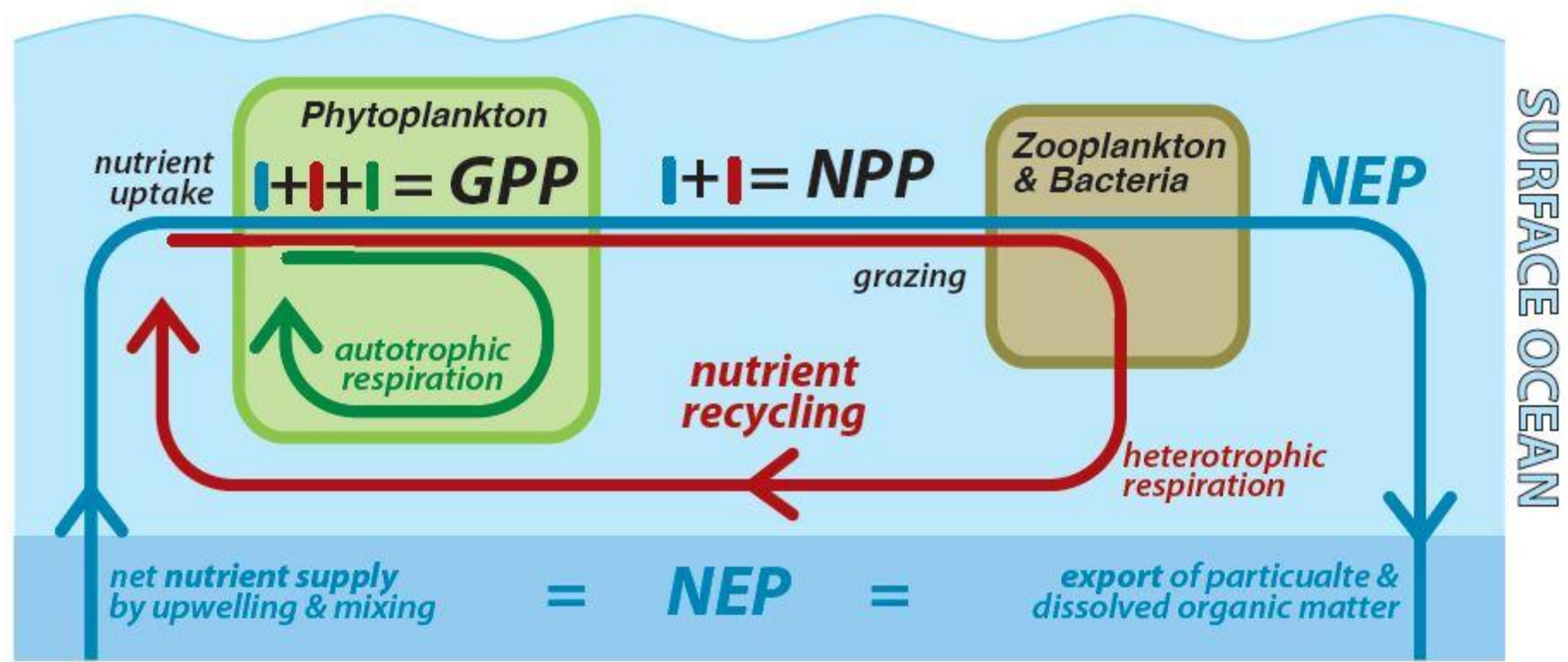
IPSL-CM4 /  
PISCES model



# Photosynthesis



- ✓ Primary Productivity is the rate at which light energy or inorganic chemical energy is converted to the chemical energy of organic compounds by autotrophs in an ecosystem.
- ✓ In the ocean, photosynthesis is performed by phytoplankton in the sunlit or euphotic zone.
- ✓ This process takes carbon dioxide and water and combines them with the help of the energy contained in sunlight creates a monosaccharide and oxygen

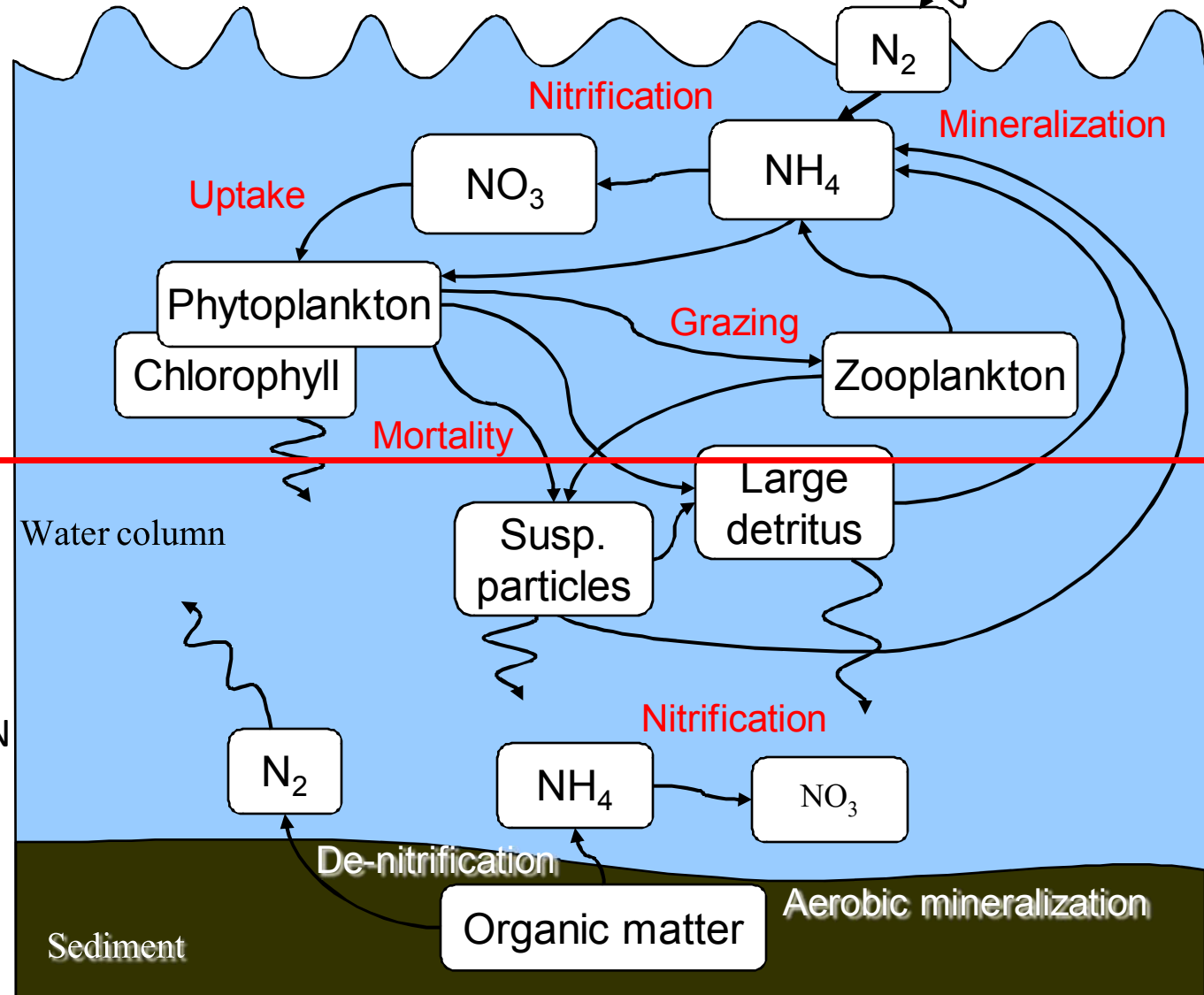


- ✓ “Gross primary production” (GPP) refers to the total rate of organic carbon production by autotrophs
- ✓ “respiration” refers to the energy-yielding oxidation of organic carbon back to carbon dioxide.
- ✓ “Net primary production” (NPP) is GPP minus the autotrophs’ own rate of respiration; it is thus the rate at which the full metabo-lism of phytoplankton produces biomass.



Mix Layer  
depth

Continental shelf  
sediments are  
responsible for up  
to 67% of marine N  
denitrification  
estimates



De-nitrification – the removal of fixed N, mostly  $NO_3^-$ , resulting in the formation of non-biologically available N, primarily  $N_2$  gas



# Partition of Anthropogenic Carbon Emissions into Sinks [2000-2006]



45% of all  $\text{CO}_2$  emissions accumulated  
in the atmosphere



55% were removed by natural sinks

Ocean removes ~ 24%



Land removes ~ 30%



## I. Wavelength-resolved models (WRMs)

$$\sum PP = \int_{\lambda=400}^{700} \int_{t=\text{sunrise}}^{\text{sunset}} \int_{z=0}^{Z_{eu}} \Phi(\lambda, t, z) \times \text{PAR}(\lambda, t, z) \times \text{Chl}(z) d\lambda dt dz - R$$

- ✓ WRM convert absorbed radiation i.e. Photosynthetically Utilizable Radiation (PUR) into net photosynthesis using a suit of empirical quantum efficiency models based on photosynthesis-irradiance variables

## II. Wavelength-integrated models (WIMs)

$$\sum PP = \int_{t=\text{sunrise}}^{\text{sunset}} \int_{z=0}^{Z_{eu}} \varphi(t, z) \times \text{PAR}(t, z) \times \text{Chl}(z) dt dz - R$$

- ✓ WIM eliminated wavelength dependencies and NPP is described as a function of PAR rather than PUR and calculated by integrating PAR dependent photosynthesis-irradiance function over depth and time





## II. Time-integrated models (TIMs)

$$\sum PP = \int_{z=0}^{z_{eu}} P^b(z) \times PAR(z) \times DL \times Chl(z) dz$$

- ✓ TIM eliminates time-dependent resolution in solar irradiance. TIM intrinsically integrate a range of photosynthetic rates into a single productivity value.

## IV. Depth-integrated models (DIMs)

$$\sum PP = P^b_{opt} \times f[PAR(0)] \times DL \times Chl \times Z_{eu}$$

- ✓ DIM includes all models lacking any explicit description of the vertically resolved component found in TIM, WRM and WIM.



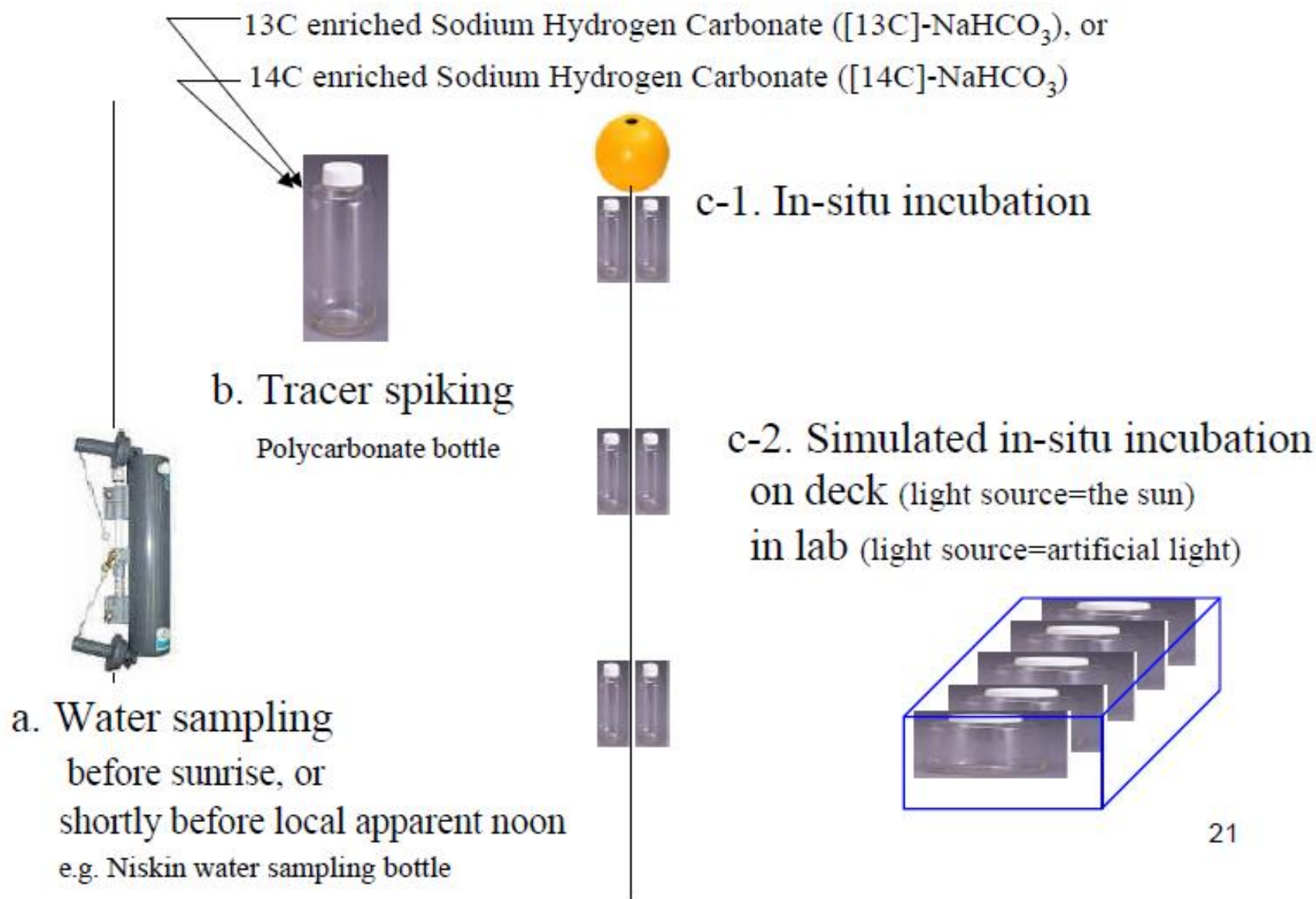
## 1) Light/Dark Bottle Method

- Calculate DO in both bottles at  $t=t_0$ ,  $t=t_1$
- $GPP = (DO_{t_1}^{light} - DO_{t_1}^{dark}) / (t_1 - t_0)$
- $NPP = (DO_{t_1}^{light} - DO_{t_0}^{light}) / (t_1 - t_0)$
- Quick but only an estimation (based on  $O_2$  consumption)

## 2) Nitrogen/Carbon isotop method

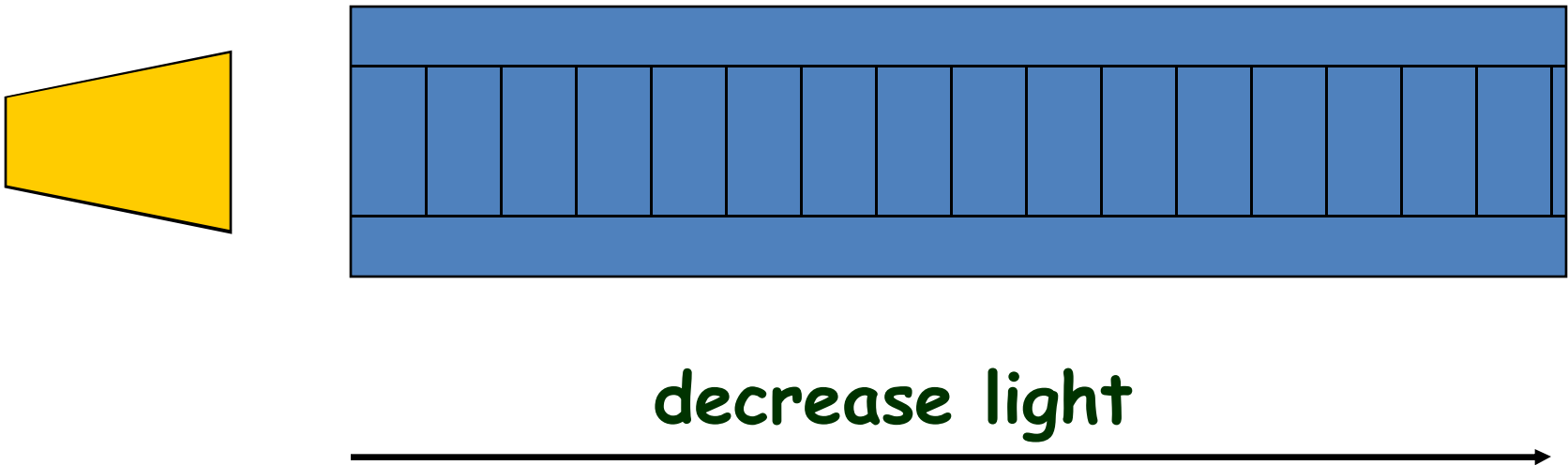
- Inoculate  $C^{13}/C^{14}$  (DIC), various light-levels, temp. regulation
- Filter samples, flush excess isotops, POC retained on filter paper
- Measure ratio to  $C^{12}$  with IRMS/Mw-IRMS, highly precise



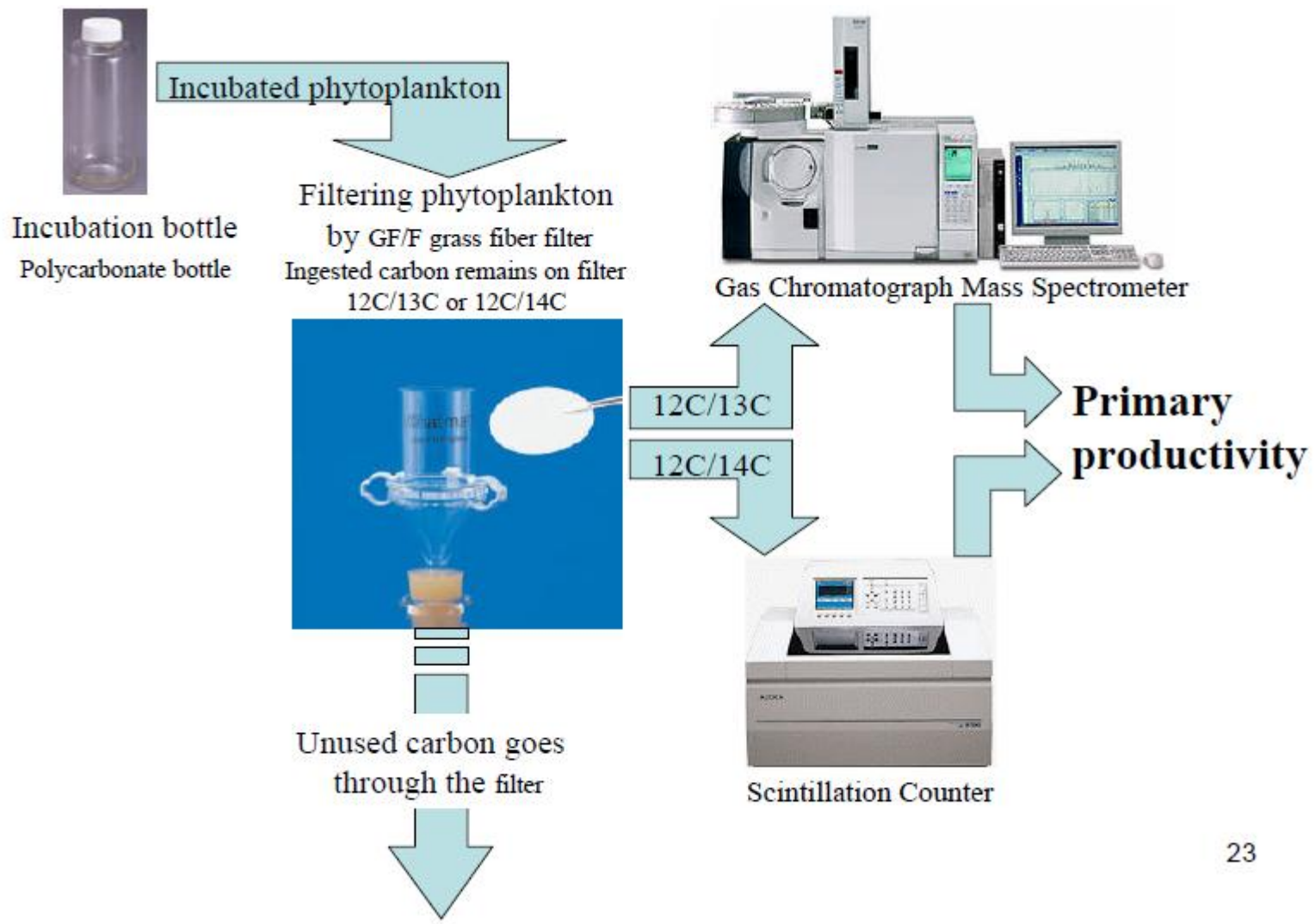




# light gradient Incubator







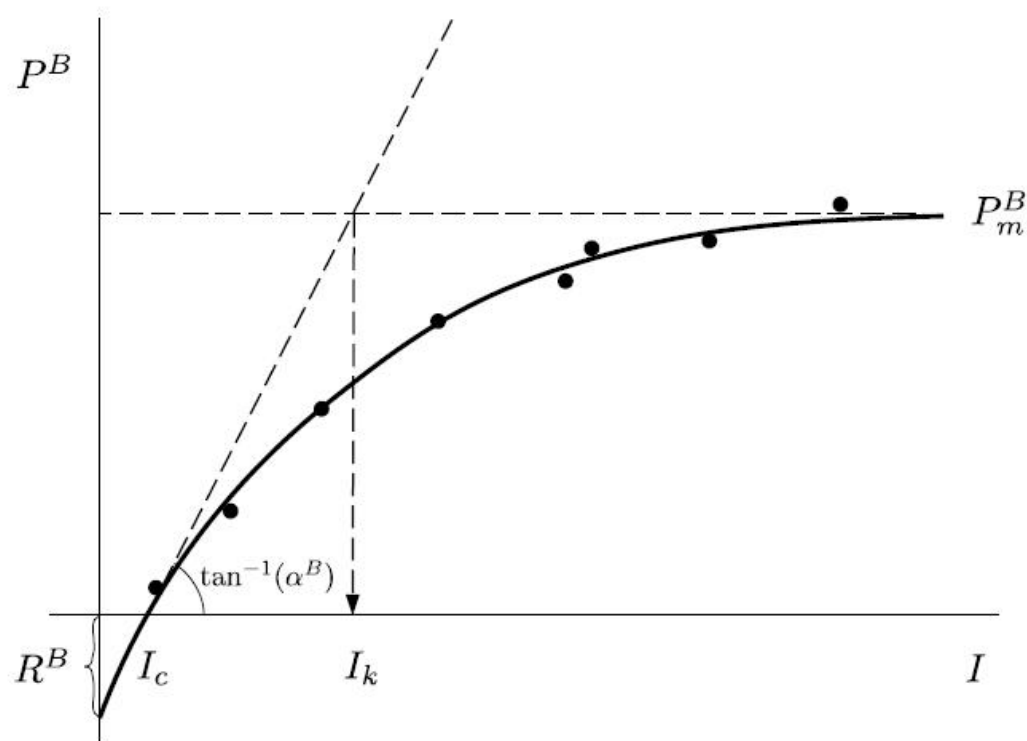


Figure 1. The photosynthesis – light curve (solid line) as fitted to imaginary experimental data (solid circles). The broken lines are construction lines to show the meaning of the parameters.

$$R^B = 0, \text{ we have } P_m^B / I_k = \alpha^B, \quad I_k = P_m^B / \alpha^B$$

$$I_* = I / I_k = \alpha^B I / P_m^B$$



$$P^B(I) = p^B(I; \alpha^B, P_m^B),$$

Table 1. Some commonly-used, two-parameter representations of the  $P^B$  vs  $I$  curve, with references to their first appearance in the phytoplankton literature. The function  $p^B$  is defined by the equation  $P^B = p^B(I; \alpha^B, P^B)$ . Also given, in the third column, is the equivalent function expressed in terms of the dimensionless irradiance,  $I_* = I/I_k$ , where  $I_k = P_m^B/\alpha^B$ .

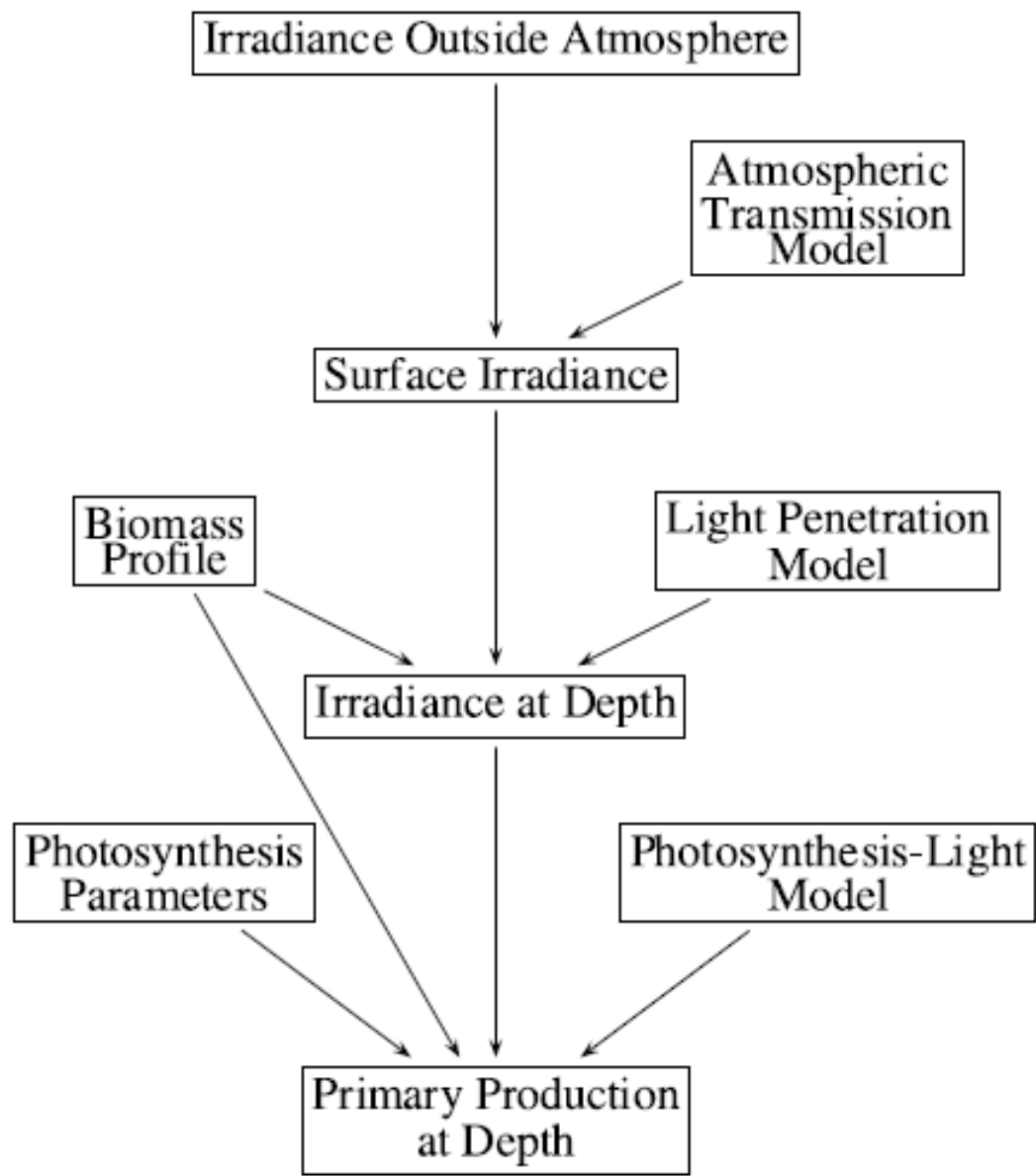
Reference	$p^B(I)$	$p^B(I_*)$
Blackman 1905	$\alpha^B I$ for $I \leq I_k$ ; $P_m^B$ otherwise	$P_m^B I_*$ for $I_* \leq 1$ ; $P_m^B$ otherwise
Smith 1936	$\frac{\alpha^B I}{\sqrt{1 + (\alpha^B I/P_m^B)^2}}$	$\frac{P_m^B I_*}{\sqrt{1 + I_*^2}}$
Tamiya 1951 †	$\frac{P_m^B \alpha^B I}{P_m^B + \alpha^B I}$	$\frac{P_m^B I_*}{1 + I_*}$
Jassby & Platt 1976	$P_m^B \tanh(\alpha^B I/P_m^B)$	$P_m^B \tanh(I_*)$
Platt <i>et al.</i> 1980 ¶	$P_m^B (1 - \exp(-\alpha^B I/P_m^B))$	$P_m^B (1 - \exp(-I_*))$





Table 1. Some commonly-used, two-parameter representations of the  $P^B$  vs  $I$  curve, with references to their first appearance in the phytoplankton literature. The function  $p^B$  is defined by the equation  $P^B = p^B(I; \alpha^B, P_m^B)$ . The equations are given in their non-spectral form,  $p^B(I)$ , and their spectral equivalents  $p^B(I(\lambda))$ .

Reference	$p^B(I)$	$p^B(I(\lambda))$
Blackman 1905	$\begin{cases} \alpha^B I, & \text{for } I \leq I_k; \\ P_m^B, & \text{for } I > I_k. \end{cases}$	$\begin{cases} \int \alpha^B(\lambda) I(\lambda) d\lambda, & \text{for } I \leq I_{k,\lambda}; \\ P_m^B, & \text{for } I > I_{k,\lambda}. \end{cases}$
Smith 1936	$\frac{\alpha^B I}{\sqrt{1 + (\alpha^B I / P_m^B)^2}}$	$\frac{\int \alpha^B(\lambda) I(\lambda) d\lambda}{\sqrt{1 + [(1/P_m^B)(\int \alpha^B(\lambda) I(\lambda) d\lambda)]^2}}$
Tamiya 1951 †	$\frac{P_m^B \alpha^B I}{P_m^B + \alpha^B I}$	$\frac{P_m^B \int \alpha^B(\lambda) I(\lambda) d\lambda}{P_m^B + \int \alpha^B(\lambda) I(\lambda) d\lambda}$
Jassby & Platt 1976	$P_m^B \tanh(\alpha^B I / P_m^B)$	$P_m^B \tanh\left((1/P_m^B) \int \alpha^B(\lambda) I(\lambda) d\lambda\right)$
Platt <i>et al.</i> 1980 ¶	$P_m^B (1 - \exp(-\alpha^B I / P_m^B))$	$P_m^B \left(1 - \exp\left(-(1/P_m^B) \int \alpha^B(\lambda) I(\lambda) d\lambda\right)\right)$



# PP modeling

Table 1. Some commonly-used, two-parameter representations of the  $P^B$  vs  $I$  curve, with references to their first appearance in the phytoplankton literature. The function  $P^B$  is defined by the equation  $P^B = p^B(I; \alpha^B, P_m^B)$ . Also given, in the third column, is the equivalent function expressed in terms of the dimensionless irradiance,  $I_* = I/I_k$ , where  $I_k = P_m^B/\alpha^B$ .



Reference	$p^B(I)$	$p^B(I_*)$
Blackman 1905	$\alpha^B I$ for $I < I_k$ ; $P_m^B$ otherwise	$P_m^B I_*$ for $I_* < 1$ ; $P_m^B$ otherwise
Smith 1936	$\frac{\alpha^B I}{\sqrt{1 + (\alpha^B I/P_m^B)^2}}$	$\frac{P_m^B I_*}{\sqrt{1 + I_*^2}}$
Tamiya 1951 †	$\frac{I^B \alpha^B I}{P_m^B + \alpha^B I}$	$\frac{I^B I_*}{1 + I_*}$
Jassby & Platt 1976	$P_m^B \tanh(\alpha^B I/P_m^B)$	$P_m^B \tanh(I_*)$
Platt <i>et al.</i> 1980 ¶	$P_m^B (1 - \exp(-\alpha^B I/P_m^B))$	$P_m^B (1 - \exp(-I_*))$

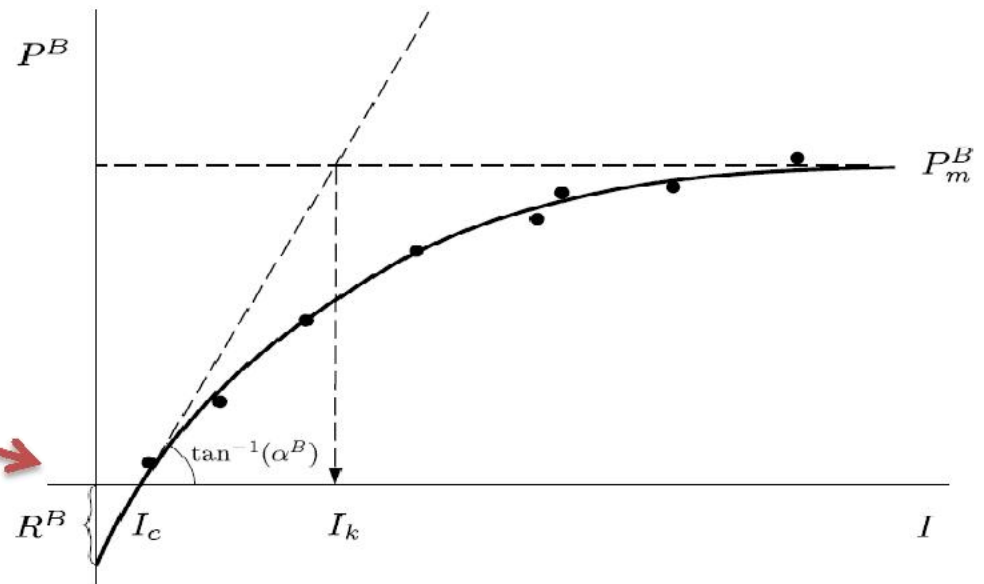
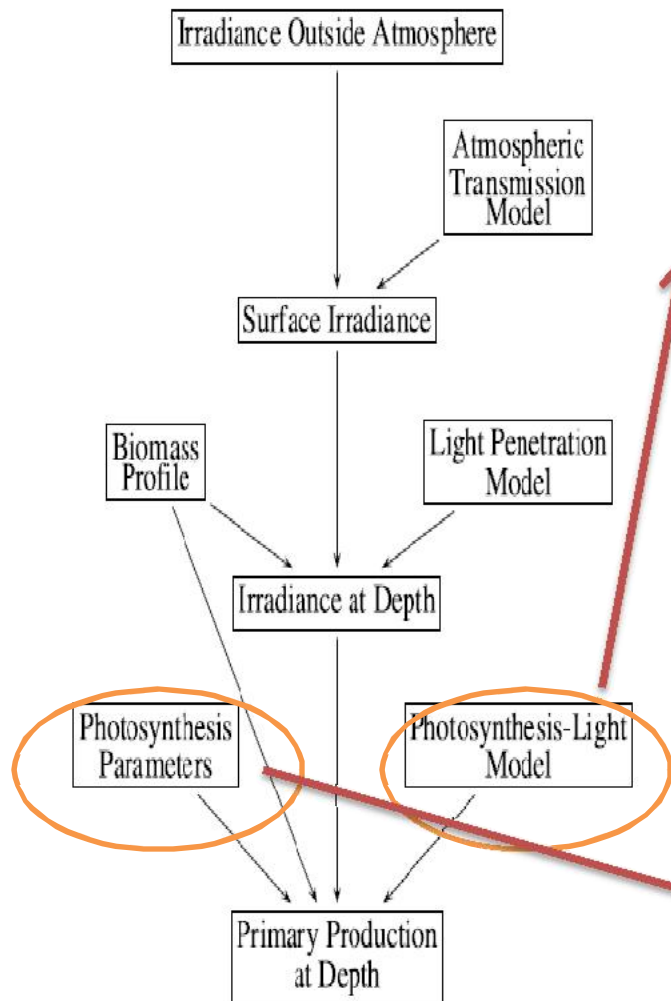
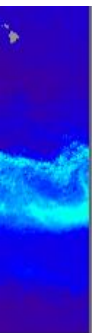


Figure 1. A flow chart describing the steps involved in computation of primary production at depth in the water column, using a light-dependent model of primary production. A number of possibilities exist for the execution of each of the steps involved in the calculation, and the success of the venture will depend on careful selection of suitable protocols for each of the steps.

Figure 1. The photosynthesis – light curve (solid line) as fitted to imaginary experimental (solid circles). The broken lines are construction lines to show the meaning of the parameters.





# PRODUCTIVITY

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## Welcome to the *Ocean Productivity* Home Page

The diversity of life on Earth is astonishing, yet most of the ecosystems you and I are familiar with share a common dependence on a miraculous process called *photosynthesis*. Photosynthesis uses the energy in sunlight to fix carbon dioxide (CO<sub>2</sub>) into organic material. Aquatic and terrestrial photosynthetic plants use some of their newly formed carbon products immediately for energy and maintenance. The remaining photosynthetic products are available for plant growth or consumption by the heterotrophic community. We refer to this "available" carbon as *net primary production*, and it is equal to gross photosynthetic carbon fixation minus the carbon respired to support maintenance requirements of the whole plant.

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A consumer's guide to phytoplankton primary productivity models

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nature Education

## The Biological Productivity of the Ocean

Daniel M. Sigman<sup>1</sup> & Mathis P. Hain<sup>1,2</sup> © 2012 Nature Education

Productivity fuels life in the ocean, drives its chemical cycles, and lowers atmospheric carbon dioxide. Nutrient uptake and export interact with circulation to yield distinct ocean regimes.



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Operational estimation of primary production at large geographical scales

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## Estimation of Marine Primary Productivity From Satellite-Derived Phytoplankton Absorption Data

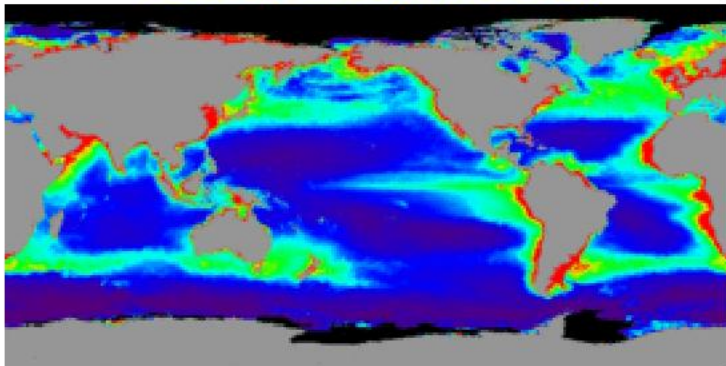
Sheng Ma, Zui Tao, Xiaofeng Yang, Member, IEEE, Yang Yu, Xuan Zhou, Wentao Ma, and Ziwei Li

Discovery and Use of Operational Ocean Data Products and Services, June 2018

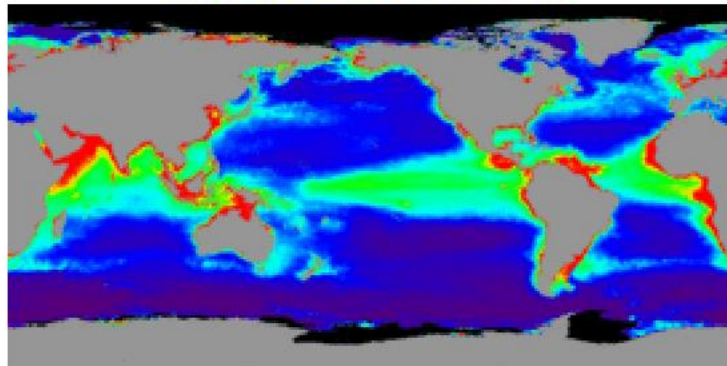


## Annual Net Primary Production for 2003

VGPM (chlorophyll based)



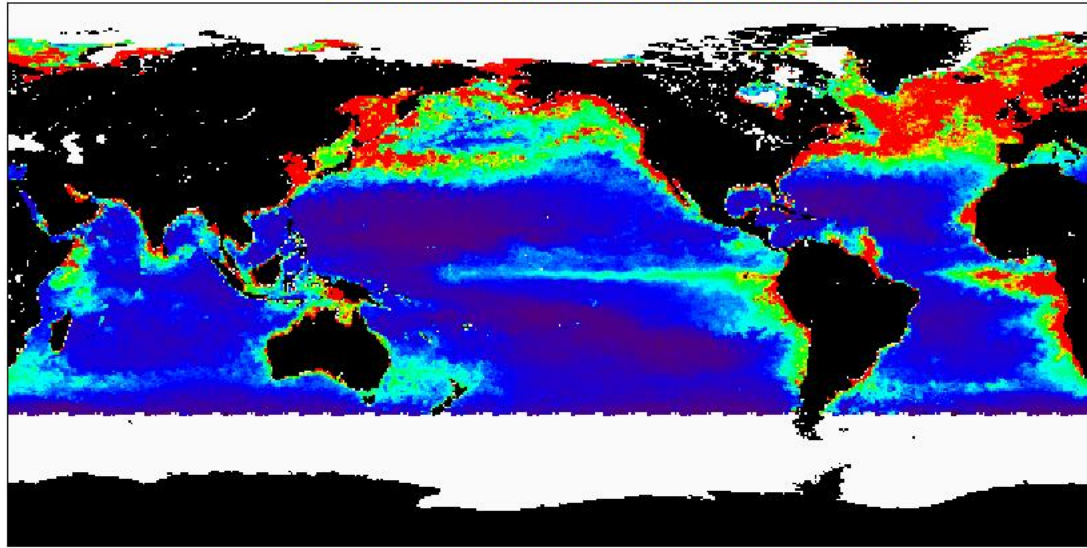
Eppley (VGPM variant)



$pb_{opt}$  = *polynomial* function of SST

$pb_{opt}$  = *exponential* function of SST

## Monthly Net Primary Production: June , 2005





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